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<p>(21) International Application Number: PCT/US97/07688</p> <p>(22) International Filing Date: 1 May 1997 (01.05.97)</p> <p>(30) Priority Data: 60/016,345 1 May 1996 (01.05.96) US</p> <p>(60) Parent Application or Grant (63) Related by Continuation US 60/016,345 (CIP) Filed on 1 May 1996 (01.05.96)</p> <p>(71) Applicant (for all designated States except US): PIONEER HIL-BRED INTERNATIONAL, INC. [US/US]; Darwin Build-ing, 7100 N.W. 62nd Avenue, P.O. Box 1000, Johnston, IA 50131 (US).</p> <p>(72) Inventors; and (75) Inventors/Applicants (for US only): GORDON-KAMM, William [US/US]; 3916 67th Street, Urbandale, IA 50322 (US). PIERCE, Dorothy, A. [US/US]; 9124 Hammontree Drive, Urbandale, IA 50322 (US). BOWEN, Benjamin [GB/US]; 3008 36th Street, Des Moines, IA 50310 (US). BIDNEY, Dennis [US/US]; 8385 Plum Drive, Urbandale, IA 50322 (US). ROSS, Margit [US/US]; 5534 Kensington</p>	<p>Circle, Johnston, IA 50131 (US). SCHELONGE, Christopher [US/US]; 5314 Ovid Avenue, Des Moines, IA 50310 (US). MILLER, Mike [US/US]; 59 W. Buchanan Street, Winterset, IA 50273 (US). SANDAHL, Gary [US/US]; 1649 Thornwood Road, West Des Moines, IA 50265 (US). WANG, Lijuan [CN/US]; 4728 80th Place, Urbandale, IA 50322 (US).</p> <p>(74) Agents: BENT, Stephen, A. et al.; Foley & Lardner, Suite 500, 3000 K Street, N.W., Washington, DC 20007-5109 (US).</p> <p>(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU. ARIPO patent (GH, KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).</p> <p>Published Without international search report and to be republished upon receipt of that report.</p>	
<p>(54) Title: USE OF THE GREEN FLUORESCENT PROTEIN AS A SCREENABLE MARKER FOR PLANT TRANSFORMATION</p> <p>(57) Abstract</p> <p>A method for the production of transgenic plants is provided in which a vector carrying a gene encoding the green fluorescent protein is introduced into cells, the cells are screened for the protein and transformed cells are selected and regenerated. The cellular toxicity of the green fluorescent protein is circumvented by regulating expression of the gene encoding the protein or directing the protein to a subcellular compartment where it is not toxic to the cell. DNA constructs are provided for cell transformation in which the expression of a gene encoding the green fluorescent protein is placed under the control of an inducible promoter. In addition, DNA constructs are provided in which a nucleotide sequence encoding the green fluorescent protein is operably linked to a signal sequence which directs the expressed protein to a subcellular compartment where the protein is not toxic to the cell. Oxidative stress to plant cells transformed with GFP also can be ameliorated by transforming cells with an expression vector comprising genes encoding GFP and an oxygen scavenger enzyme such as superoxide dismutase. The toxicity of GFP in transformed plants can be eliminated by excising the screenable marker gene following detection of transformed cells or sectors. The FLP/FRT system is used in conjunction with GFP as a visible marker for transformation and FRT excision. A nucleotide sequence optimized for expression of the green fluorescent protein in plants is also provided.</p>		

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USE OF THE GREEN FLUORESCENT PROTEIN AS A SCREENABLE
MARKER FOR PLANT TRANSFORMATION

BACKGROUND OF THE INVENTION

1. Field of the Invention

5 The present invention relates to a method of plant transformation in which a DNA construct carrying a gene encoding the green fluorescent protein (GFP) is introduced into plant cells which are then screened for the presence of the protein and transformed cells are regenerated into transgenic plants. In particular, the present invention provides methods for circumventing the cellular toxicity of the GFP by regulating expression of the gene encoding the protein or directing the protein to a subcellular compartment where it is not toxic to the cell. The present invention provides DNA constructs for cell transformation in which expression of a gene encoding the GFP is placed under the control of an inducible, constitutive or tissue-specific promoter. In addition, DNA constructs are provided in which a nucleotide sequence encoding the GFP is operably linked to a signal or targeting sequence which directs the expressed protein to a subcellular compartment where the protein is not toxic to the cell. Moreover, the present invention provides a nucleotide sequence encoding GFP that is optimized for expression of the GFP gene in

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plants and to GFP-encoding nucleotide sequences that code for light-shifted versions of GFP. The present invention also provides a method for selecting plant cells transformed with a gene encoding a screenable marker flanked on the 5-prime and 3-prime ends with a recombinase-specific target sequence, and introducing a gene encoding a site specific recombinase into the transformed plant cells and selecting transformed plant cells that no longer express the screenable marker. In addition, the present invention provides a method of reducing GFP toxicity by transforming plant cells with a gene encoding the GFP together with a gene encoding an oxygen scavenger such as superoxidase dismutase.

2. Background

Expression vectors include at least one genetic marker that allows transformed cells to be either recovered by negative selection, i.e. inhibiting growth of cells that do not contain the selectable marker gene, or by screening for product encoded by the genetic marker. Many of the commonly used selectable marker genes for plant transformation were isolated from bacteria and code for enzymes that metabolically detoxify a selective chemical agent which may be an antibiotic or a herbicide. Other selectable marker genes encode an altered target which is insensitive to the inhibitor.

The most commonly used selectable marker gene for plant transformation is the neomycin phosphotransferase II (*nptII*) gene, isolated from *Tn5*, which when placed under the control of plant regulatory signals confers resistance to kanamycin. Fraley et al., *Proc. Natl. Acad. Sci. U.S.A.*, 80: 4803 (1983). Another commonly used selectable marker gene is the hygromycin phosphotransferase gene which confers resistance to the antibiotic hygromycin. Vanden Elzen et al., *Plant Mol. Biol.*, 5: 299 (1985).

Additional selectable marker genes of bacterial origin that confer resistance to antibiotics include gentamycin acetyl transferase, streptomycin phosphotransferase, aminoglycoside-3'-adenyl transferase, the bleomycin resistance determinant. Hayford et al., *Plant Physiol.* 86: 1216 (1988), Jones et al., *Mol. Gen. Genet.*, 210: 86 (1987), Svab et al., *Plant Mol. Biol.* 14: 197 (1990), Hille et al., *Plant Mol. Biol.* 7: 171 (1986). Other selectable marker genes confer resistance to herbicides such as glyphosate, glufosinate or broxynil. Comai et al., *Nature* 317: 741-744 (1985), Gordon-Kamm et al., *Plant Cell* 2: 603-618 (1990) and Stalker et al., *Science* 242: 419-423 (1988).

Other selectable marker genes for plant transformation are not of bacterial origin. These genes include, for example, mouse dihydrofolate reductase, plant 5-enolpyruvylshikimate-3-phosphate synthase and plant acetolactate synthase. Eichholtz et al., *Somatic Cell Mol. Genet.* 13: 67 (1987), Shah et al., *Science* 233: 478 (1986), Charest et al., *Plant Cell Rep.* 8: 643 (1990).

Although many of these markers have been used for selecting transformed plant tissue, these selection systems involving toxic chemical agents can have disadvantages or limitations. One disadvantage is that it may be difficult to recover normal, viable transformed plants directly from chemical selection. Everett et al., *Bio/Technology* 5: 1201-1204 (1987). Another disadvantage is that not all selectable marker systems work for all tissues, in all plant species, due in part to differences in sensitivity of a particular tissue or plant species to the selective agent. The success of any given marker for transformation of a given plant species is not easily predicted. Moreover, potential regulatory issues surrounding the use of antibiotic resistance genes and the use of herbicide resistance genes for plant species capable of outcrossing with weedy species are additional disadvantages of these markers.

Another class of marker genes for plant transformation require screening of presumptively transformed plant cells rather than direct genetic selection of transformed cells for resistance to a toxic substance such as an antibiotic. These genes are particularly useful to quantify or visualize the spatial pattern of expression of a gene in specific tissues and are frequently referred to as reporter genes because they can be fused to a gene or gene regulatory sequence for the investigation of gene expression. Commonly used genes for screening presumptively transformed cells include β -glucuronidase (GUS), β -galactosidase, luciferase, and chloramphenicol acetyltransferase. Jefferson, R.A., *Plant Mol. Biol. Rep.* 5: 387 (1987).. Teeri et al., *EMBO J.* 8: 343 (1989), Koncz et al., *Proc. Natl. Acad. Sci. U.S.A.* 84: 131 (1987), De Block et al., *EMBO J.* 3: 1681 (1984). Another approach to the identification of relatively rare transformation events has been use of a gene that encodes a dominant constitutive regulator of the *Zea mays* anthocyanin pigmentation pathway. Ludwig et al., *Science* 247: 449 (1990).

Although chemical selection of plant cells transformed with selectable marker genes has been successful with plant species and varieties that are easily cultured in vitro, the choice of selectable marker systems that have been shown to be successful for cereals and many other agronomically important plant species is very limited. In general, plant species that tend toward organogenesis and/or shoot propagation have been difficult to transform by means of chemical selection. The success rate with these plant species continues to improve, however, as evidenced by recent advances in Type I selection of maize inbreds and small grain cereals such as barley. Koziel et al., *Bio/Technology* 11: 194-200 (1993) and Mendel et al. In: *Transformation of Plants and Soil Microorganisms*, Wang et al. eds., Cambridge Press (1995).

Likewise, there has been little success in using visual screening methods for primary identification of transformed cells. The GUS gene was used to investigate germline transmission. McCabe et al., *Plant Physiol.*, 87(3): 671 (1988) and McCabe et al., *Plant Cell Tissue Organ Cult.* 33(3): 227 (1993). Histochemical staining for GUS activity was used to locate transgenic sectors in cotton and soybean transformants that ultimately produced transgenic seeds. Since histochemical analysis for GUS activity requires destruction of portions of the presumptively transformed plant tissue, this method is labor intensive and impractical for routine production of transgenic plants. This method is particularly unsuitable for plant species such as maize and other cereals in which transformants are recovered, even under optimum conditions, at low frequency. Recovery of transformed progeny was reported once in barley using GUS expression as a screening tool, but the method was found to be very labor-intensive. Ritala et al., *Plant Mol. Biol.* 24: 317-325 (1994). There have been no reports at all of success with GUS or other screenable markers with maize.

More recently, *in vivo* methods for visualizing GUS activity that do not require destruction of plant tissue have been made available. Molecular Probes Publication 2908, *Imagene Green™*, p. 1-4 (1993) and Naleway et al., *J. Cell Biol.* 115: 151a (1991). However, these *in vivo* methods for visualizing GUS activity have not proven useful for recovery of transformed cells because of low sensitivity and high fluorescent backgrounds.

Despite the fact that luciferase genes have been available for many years, this strategy for visualizing transformed cells has also not been successfully adapted for routine recovery of plant transformants. The luciferase-based screening methods are limited by the fact that most of these systems require the presence of luciferin, a compatible luciferase and an exogenously supplied cofactor. In the absence of the substrate,

enzyme or cofactor, the system does not bioluminesce. Cells transformed with a luciferase gene must have cell walls and plasma membranes that are permeable, or rendered permeable, to a compatible luciferin in order to detect bioluminescence. For example, tobacco plants regenerated from cells transformed with a firefly luciferase gene and exposed to a liquid medium containing firefly luciferin exhibited bioluminescence primarily along their major veins. Ow et al., *Science* 234: 856 (1986). Accordingly, a screening method has not been successfully developed for routine plant transformation that does not involve chemical selection or assays, often labor-intensive, that require the sacrifice or destruction of tissue samples for analysis.

A gene encoding GFP has been utilized as a marker for gene expression in prokaryotic and eukaryotic cells. Chalfie et al., *Science* 263: 802 (1994). Many cnidarians utilize GFPs as energy-transfer acceptors in bioluminescence. A gene encoding GFP isolated from a cnidarian and expressed in a heterologous prokaryotic or eukaryotic host produces a protein capable of fluorescence. A cDNA encoding the *Aequorea victoria* GFP produced a fluorescent product when expressed in *Escherichia coli* or *Caenorhabditis elegans* cells. Green fluorescence was detected in transformed cells upon illumination with a long-wave ultraviolet (UV) source without having to supply substrates or cofactors. In addition, fluorescence was stable for at least 10 min when illuminated with 450 to 490 nm light. Transformation of plant cells with a gene encoding GFP and detection of fluorescence has been reported. Haseloff et al., *TIG* 11: 328-329 (1995). Transformed *Arabidopsis* cells could be regenerated into whole plants. However, the regenerated plants expressing GFP exhibited signs of mild to moderate toxicity in the light compared to plants not expressing GFP. The strongest GFP expressors proved more difficult to regenerate. Likewise, in a recent report describing GFP expression in

kanamycin-selected tobacco transformants, Chiu et al.,
Current Biol. 6: 325-330 (1996) note that high GFP
expression levels inhibited regeneration of transgenic
plants.

5 A need therefore exists for a cell transformation
method which does not rely, or does not rely solely on,
selection of cells carrying a gene that confers
resistance to a toxic substance. A need exists for a
method for efficiently and easily identifying transformed
10 plant cells with a visual screenable marker. A need also
exists for a method of cell transformation that does not
require destruction of presumptively transformed tissue
to assay for the presence of a selectable marker gene.
A need exists for a method for cell transformation that
15 combines a selectable marker gene and a screenable marker
gene. In addition, a need exists for a method of cell
transformation which does not require exogenous supply of
a substrate or cofactor for detection of the polypeptide
encoded by a selectable marker gene. Yet another need
20 exists for a method of cell transformation that
circumvents the cellular toxicity of the GFP.

SUMMARY OF THE INVENTION

25 Accordingly, it is an object of the present
invention to provide a method for cell transformation
which does not depend on selection of cells carrying a
gene that confers resistance to a toxic substance.

30 It is another object of the present invention to
provide a method for cell transformation which combines
the selection of cells carrying a gene that confers
resistance to a toxic substance with screening cells for
the presence of a substance that renders transformed
cells identifiable.

35 It is a further object of the present invention to
provide a method for cell transformation which does not
require destruction of presumptively transformed tissue
to assay for the presence of a selectable marker gene.

Yet another object of the present invention is to provide a method for cell transformation which does not require that an exogenous substrate or cofactor be provided to assay for a polypeptide encoded by a selectable marker gene.

It is another object of the present invention to provide a method for cell transformation that circumvents the cellular toxicity of the GFP.

These and other objects are achieved, in accordance with one embodiment of the present invention, by providing an isolated DNA molecule comprising a nucleotide sequence encoding the GFP operably linked to an inducible promoter. The inducible promoter can be selected from the group consisting of the estrogen-inducible promoter, the estradiol-inducible promoter, the ACE1 promoter, the IN2 promoter and the tetracycline repressor promoter.

Also provided is an isolated DNA molecule comprising as nucleotide sequence encoding the GFP wherein the nucleotide sequence is operably linked to a targeting sequence for subcellular localization which directs a protein to a subcellular compartment.

An isolated DNA molecule is provided comprising a nucleotide sequence selected from the group consisting of (a) SEQ ID NO: 1; (b) a nucleotide sequence that has substantial sequence similarity with SEQ ID NO: 1; and (c) a functional fragment of (a) or (b), wherein said DNA molecule encodes a GFP. The nucleotide sequence encoding a GFP may be operably linked to a nucleotide sequence encoding a targeting sequence for subcellular localization which directs a protein to a subcellular compartment. The nucleotide sequence encoding a GFP and a targeting sequence may further comprise a promoter operably linked to said nucleotide sequence.

Also provided are expression vectors comprising DNA molecules encoding a GFP, optionally operably linked to an inducible promoter and/or targeting sequence. The expression vector of the instant invention may carry a

nucleotide sequence encoding a foreign protein operably linked to a second promoter.

Also provided is a method of using the expression vectors of the instant invention to produce a transformed plant, comprising the steps of introducing an expression carrying a gene encoding GFP into regenerable plant cells and selecting cells containing the GFP for regeneration. The method of the instant invention may include the step of inducing GFP expression where the nucleotide sequence encoding GFP is operably linked to an inducible promoter. The regenerable plant cells utilized in the method of the instant invention are selected from the group consisting of *Zea*, *Brassica* and *Helianthus* cells.

Also provided are transgenic plants expressing the isolated DNA molecule encoding GFP. In addition, transgenic plants comprising a vector carrying a nucleotide sequence encoding GFP are provided.

A method for producing a transgenic plant is provided comprising the steps of (a) constructing an expression vector comprising (i) a first promoter which is an inducible promoter, operably linked to a nucleotide sequence encoding a GFP, and (ii) a second promoter operably linked to a foreign gene; (b) introducing said expression vector into regenerable plant cells; (c) inducing expression of the gene encoding the GFP and selecting transformed plant cells containing said protein; and (d) regenerating transformed plants from said selected transformed plant cells. The inducible promoter may be selected from the group including the estrogen-inducible promoter, the estradiol-inducible promoter, the ACE1 promoter, the IN2 promoter and the tetracycline repressor promoter.

Also provided is a method for producing a transgenic plant, comprising the steps: (a) constructing an expression vector comprising (i) a first promoter operably linked to a nucleotide sequence encoding a sequence for subcellular localization which directs a protein to a subcellular compartment which is operably

linked to a nucleotide sequence encoding a GFP, and (ii) a second promoter operably linked to a foreign gene; (b) introducing said expression vector into regenerable plant cells; (c) selecting transformed plant cells containing said GFP; and (d) regenerating transformed plants from said selected transformed plant cells. The targeting sequence for subcellular localization directs the GFP to the mitochondria, chloroplasts, peroxisomes, vacuole, endoplasmic reticulum, cell wall or for secretion generally into the apoplast. This method may include a nucleotide sequence encoding a GFP selected from the group consisting of: (a) SEQ ID NO: 1; (b) a nucleotide sequence that has substantial sequence similarity with SEQ ID NO: 1; and (c) a functional fragment of (a) or (b).

Also provided is a method for producing a transgenic plant, comprising the steps: (a) constructing an expression vector comprising (i) a first promoter operably linked to a nucleotide sequence encoding a screenable marker flanked on the 5-prime and 3-prime ends with a recombinase-specific target sequence, and (ii) a second promoter operably linked to a foreign gene; (b) introducing said expression vector into regenerable plant cells; (c) selecting transformed plant cells containing said screenable marker; (d) transforming the plant cells with a second expression vector containing a gene encoding a site-specific recombinase; (e) selecting plant cells that no longer express the screenable marker; (f) regenerating transformed plants from said selected transformed plant cells; and (g) isolating said foreign protein. The site-specific recombinase may be selected from the group consisting of the FLP/FRT, Ac/DS and cre/lox systems. In addition, the screenable marker may be the GFP.

An isolated DNA molecule comprising a nucleotide sequence encoding the GFP fused in frame to a nucleotide sequence encoding superoxide dismutase is also provided. Cells transformed with the DNA molecule contain a fusion

protein that (i) produces green fluorescence in the presence of UV to blue light and (ii) displays superoxide dismutase activity.

Also provided is a method for producing a transgenic plant comprising: (a) constructing an expression vector comprising (i) a first promoter operably linked to a nucleotide sequence encoding a GFP, and (ii) a second promoter operably linked to a gene encoding an enzyme that is an oxygen scavenger, and (iii) a third promoter operably linked to a foreign gene; (b) introducing said expression vector into regenerable plant cells; (c) selecting transformed plant cells containing said GFP; and (d) regenerating transformed plants from said selected transformed plant cells. The oxygen scavenger enzyme may be superoxide dismutase.

Alternatively, a method for producing a transgenic plant is provided comprising: (a) constructing an expression vector comprising (i) a first promoter operably linked to a nucleotide sequence encoding a fusion protein comprising the GFP and an oxygen scavenger enzyme fused in frame, and (ii) a second promoter operably linked to a foreign gene; (b) introducing said expression vector into regenerable plant cells; (c) selecting transformed plant cells containing said the GFP and oxygen scavenger enzyme activity; and (d) regenerating transformed plants from said selected transformed plant cells. The oxygen scavenger enzyme may be superoxide dismutase.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 presents the nucleotide sequence [SEQ ID NO: 1] encoding a GFP with its corresponding amino acid sequence [SEQ ID NO: 2].

Figure 2 shows the Plasmid Map of PHP167.

Figure 3 shows the Plasmid Map of PHP762.

Figure 4 shows the Plasmid Map of PHP7921.

Figure 5 shows the Plasmid Map of PHP8144.

DETAILED DESCRIPTION

1. Definitions

In the description that follows, a number of terms are used extensively. The following definitions are provided to facilitate understanding of the invention.

A structural gene is a DNA sequence that is transcribed into messenger RNA (mRNA) which is then translated into a sequence of amino acids characteristic of a specific polypeptide.

A promoter is a DNA sequence that directs the transcription of a structural gene. Typically, a promoter is located in the 5-prime region of a gene, proximal to the transcriptional start site of a structural gene. If a promoter is an inducible promoter, then the rate of transcription increases in response to an inducing agent. In contrast, the rate of transcription is not regulated by an inducing agent if the promoter is a constitutive promoter. For example, a promoter may be regulated in a tissue-specific or tissue-preferred manner such that it is only active in transcribing the associated coding region in a specific tissue type(s) such as leaves, roots or meristem.

A cytoplasmically localized protein is a protein coded for by a gene which does not include any specific signal for targeting of the protein into any subcellular organelle or compartment or any signals for secretion of the protein. For example, the GFP structural gene operably linked 5-prime to a promoter and to an appropriate 3-prime sequence encodes for a protein compartmentalized in the cytoplasm.

A signal or targeting sequence is a structural peptide domain required for targeting of a given polypeptide to a subcellular organelle, subcellular compartment or secretion from the cell. As used herein, the phrase sequence for subcellular localization is

intended to refer collectively to any form of signal, targeting or retention sequence as defined herein. Signal sequences for a polypeptide directed to the chloroplast or mitochondrion are largely localized to the amino-terminus of the polypeptide. Signal sequences for a polypeptide directed to the glyoxysomes and peroxisomes are largely localized to the carboxy-terminal domains of the polypeptide. Accordingly, targeted transport of the GFP protein is accomplished by means of chemically joining in proper reading frame, or operably linking, the nucleotide sequence encoding a signal sequence to the 5-prime and/or 3-prime region of the GFP structural gene.

A mitochondrial targeting sequence facilitates the transport of a protein to a mitochondrial compartment. Typically, the mitochondrial targeting sequence is located at the amino-terminus of a polypeptide.

A secretion targeting sequence targets a polypeptide for export into the extracellular space through the ER. For example, operably linking a nucleotide sequence encoding the barley alpha amylase 1 (BAA) secretory targeting sequence to the 5-prime end of a structural gene targets the encoded protein for export into the extracellular space.

A cell wall targeting sequence targets a polypeptide for export from the cell but the polypeptide is specifically localized to the cell wall. For example, cell wall localization of a polypeptide is accomplished by operably linking a nucleotide sequence encoding BAA 5-prime, and operably linking a nucleotide sequence encoding a portion of the maize hydroxyproline-rich glycoprotein 3-prime to a gene encoding the polypeptide. Steifel et al., Plant Cell 2: 785-793 (1990).

A vacuolar signal sequence facilitates the transport of a protein to the vacuole. For example, vacuolar targeting is accomplished by fusing the BAA secretory signal sequence at the amino-terminus of the protein and a sequence encoding a vacuolar signal sequence to the carboxy-terminus. Transport of a polypeptide to the

vacuole is therefore accomplished by means operably linking a nucleotide sequence encoding BAA 5-prime, and a nucleotide sequence encoding a vacuolar signal sequence 3-prime to a gene encoding a polypeptide. Alternatively, vacuolar targeting is accomplished by constructing a nucleotide sequence comprising in the 5-prime to 3-prime direction nucleotide sequences encoding a vacuole signal sequence, BAA and a polypeptide.

An endoplasmic reticulum retention sequence targets a polypeptide for localization in the lumen of the endoplasmic reticulum. For example, a polypeptide is targeted for retention in the endoplasmic reticulum through the addition of the BAA sequence on the amino-terminus and an endoplasmic reticulum signal sequence on the carboxy-terminus of the polypeptide.

A nuclear targeting sequence facilitates transport of a polypeptide to the nucleus. Typically, the nuclear signal sequence is located at the amino-terminus of a polypeptide. In order to retain the nuclear targeted protein in the nucleus, it may be necessary to increase the molecular weight of the protein by means of fusing an unrelated protein to the carboxy-terminus of the targeted protein. For example, GFP was retained in the nucleus by operably linking a nucleotide sequence encoding a nuclear signal sequence 5-prime and a nucleotide sequence encoding maize acetolactate synthase 3-prime to a gene encoding a polypeptide.

A peroxisomal targeting sequence facilitates the transport of a polypeptide into the peroxisome. Typically, the peroxisomal signal sequence is a tripeptide located at the carboxy-terminus of a polypeptide.

A chloroplast targeting sequence facilitates the transport of a nuclear encoded protein to a chloroplast compartment. Typically, the chloroplast signal sequence is located at the amino-terminus of a polypeptide. Accordingly, transport of a polypeptide to a chloroplast compartment is accomplished by means of operably linking

the nucleotide sequence encoding a chloroplast signal sequence to the 5-prime region of a gene encoding a polypeptide.

5 An isolated DNA molecule is a fragment of DNA that is not integrated in the genomic DNA of an organism.

10 Complementary DNA (cDNA) is a single-stranded DNA molecule that is formed from an mRNA template by the enzyme reverse transcriptase. Typically, a primer complementary to portions of mRNA is employed for the initiation of reverse transcription. Those skilled in the art also use the term "cDNA" to refer to a double-stranded DNA molecule consisting of such a single-stranded DNA molecule and its complementary DNA strand.

15 The term expression refers to the biosynthesis of a gene product. For example, in the case of a structural gene, expression involves transcription of the structural gene into mRNA and the translation of mRNA into one or more polypeptides.

20 A cloning vector is a DNA molecule, such as a plasmid, cosmid, or bacteriophage, that has the capability of replicating autonomously in a host cell. Cloning vectors typically contain one or a small number of restriction endonuclease recognition sites at which foreign DNA sequences can be inserted in a determinable fashion without loss of an essential biological function of the vector, as well as a marker gene that is suitable for use in the identification and selection of cells transformed with the cloning vector. Marker genes typically include, for example, genes that provide tetracycline resistance, ampicillin resistance or
30 kanamycin resistance.

35 An expression vector is a DNA molecule comprising a gene that is expressed in a host cell. Typically, gene expression is placed under the control of certain regulatory elements, including constitutive or inducible promoters, tissue-specific regulatory elements, and enhancers. Such a gene is said to be operably linked to the regulatory elements.

Transformation includes introduction of genetic material into plant cells resulting in chromosomal integration and stable heritability through meiosis. Transformation also includes introduction of genetic material into plant cells in the form of plant viral vectors involving epichromosomal replication and gene expression which may exhibit variable properties with respect to meiotic stability.

A foreign gene refers in the present description to a DNA sequence that is operably linked to at least one heterologous regulatory element.

A recombinant host may be any prokaryotic or eukaryotic cell that contains either a cloning vector or expression vector. This term also includes those prokaryotic or eukaryotic cells that have been genetically engineered to contain the cloned gene(s) in the chromosome or genome of the host cell.

A transgenic plant is a plant having one or more plant cells that contain an expression vector.

A green fluorescent protein (GFP), or a functional fragment of a GFP, is capable of producing a green fluorescence. GFP absorbs in the UV to blue range with a peak at 395 nm and emits in the green with a peak at 510 nm. The "red-shifted" version of GFP is a modified version of GFP which absorbs between 480 to 490 nm and emits in the green with a peak at 510nm. Red-shifted GFP is abbreviated as GFP_r. The "blue fluorescent protein" is a modified version of GFP which absorbs around 380nm and emits in the blue with a peak at around 445nm. Blue fluorescent protein is abbreviated as BFP. GFP_n is a nucleotide sequence coding for GFP in which the DNA sequence has been modified based on codon preference in maize (Fig. 1).

Two nucleic acid molecules are considered to have a substantial sequence similarity if their nucleotide sequences share a similarity of at least 50%. Sequence similarity determinations can be performed, for example, using the FASTA program (Genetics Computer Group;

Madison, WI). Alternatively, sequence similarity determinations can be performed using BLAST (Basic Local Alignment Search Tool) of the Experimental GENI(F)O(R) BLAST Network Service. See Altschul et al., *J. Mol. Biol.* 215: 403 (1990). Also, see Pasternak et al., "Sequence Similarity Searches, Multiple Sequence Alignments, and Molecular Tree Building," in *METHODS IN PLANT MOLECULAR BIOLOGY AND BIOTECHNOLOGY*, Glick et al. (eds.), pages 251-267 (CRC Press 1993).

2. Reducing GFP Toxicity in Transformed Plants

Oxidative stress associated with GFP fluorescence has been suggested to cause cellular toxicity. Haseloff et al., *TIG* 11: 328-329 (1995). If GFP is to be useful as a screenable marker for plant transformation, as well as a reporter gene for detection of gene expression, strategies for reducing GFP toxicity must be available. The gene encoding GFP can be operably linked to an inducible promoter so that GFP can be transiently expressed and transformed cells identified. Alternatively, the gene encoding GFP can be operably linked to a signal sequence for targeting to an organelle or subcellular compartment or for secretion to the apoplast where GFP is not toxic to the cell.

Oxidative stress can be ameliorated by transforming cells containing GFP with an enzyme that serves as an oxygen scavenger. Such enzymes are well known in the art. For example, genes encoding the enzyme superoxide dismutase may protect against the primary oxidative stresses associated with GFP fluorescence. Balzan et al., *Proc. Nat Acad. Sci.* 92: 4219-4223 (1995). Other enzymes involved in the oxidative stress response, such as ascorbate peroxidase or glutathione S-transferase, could help mitigate secondary effects in the cell. Conklin, *Plant Physiol.* 109: 203-212 (1995).

More specifically, oxidative stress can be ameliorated by transforming cells with a DNA construct

carrying genes encoding GFP and SOD. Alternatively, the gene encoding GFP can be fused in frame to the gene encoding SOD. Cells transformed with this DNA construct produce a fusion protein that cause the cells to fluoresce in the presence of UV-blue light and express SOD activity.

The toxicity of GFP in transformed plants can be eliminated by excising the screenable marker gene following detection of transformed cells or sectors. The FLP/FRT system is used in conjunction with GFPm as a visible viable marker for FRT excision. The FLP/FRT system has been demonstrated in maize suspension cells using GUS expression as an indicator of FRT excision. Lysnik et al., NAR 21: 969-975 (1993). For example, plant cells are bombarded with a DNA construct containing the GFP gene flanked by FRT sequences as well as a foreign or agronomic gene of interest. The GFP gene may be operably linked to a constitutive promoter or an inducible promoter. In addition, the GFP gene may be operably linked to a signal sequence. Stable transformants are detected by means of screening for GFP.

Transgenic callus pieces are spread on medium and bombarded a second time with a FLP recombinase construct. Callus is monitored periodically under UV to blue illumination to detect cells that no longer express GFP. Callus pieces that no longer express GFP are regenerated and analyzed for expression of the foreign or agronomic gene. Agronomically useful transgenic plants are thereby produced that do not contain a marker gene.

The sensitivity of the screening method can be further increased by placing two markers genes between the FRT sequences. For example, plant cells are bombarded with a DNA construct containing the PAT and GFP genes flanked by FRT sequences and a foreign or agronomic gene. Stable transformants are recovered on bialaphos-containing medium and positive GFP-expression is confirmed. The transformed callus is then bombarded with FLP. The callus is then grown for 2-6 weeks with no

selection until clear GFP-null sectors can be identified. These sectors can be transferred onto bialaphos/chlorophenol red multiwell test plates to confirm bialaphos sensitivity (i.e. within 3-5 days). Callus pieces that no longer express GFP or PAT are regenerated and analyzed for expression of the foreign or agronomic gene. This permits recovery of agronomically useful transformants without any marker genes in the final product.

Likewise, the Ac/Ds system of maize can also be used in transgenic plants to excise the screenable marker gene that is transformed together with a foreign or agronomic gene. Mobilization of Ac and/or Ds has been demonstrated in diverse plants such as tomato, tobacco and *Arabidopsis*. Yoder et al., In *Tomato Technology*, Alan R. Liss, Inc. pp 189-198 (1987); Yoder et al., U.S. Patent No. 5,225,341; Baker et al., *EMBO J* 6: 1547-1554 (1987) and Lawson et al. *Mol. Gen. Genet.*, 245: 608-615 (1994). Likewise, the cre/lox recombinase system from bacteriophage P1 could also be used in conjunction with GFP. Excision of transgenes in plants using the cre/lox system was first demonstrated in tobacco. Odell et al., *Mol. Gen. Genet.*, 223: 369-378 (1990) and Dale and Ow, *Proc. Natl. Acad. Sci. USA*, 88: 10558-10562 (1991). Similar to the FLP and Ac systems described above, GFP expression provides an efficient, easily scorable phenotype for monitoring excision.

3. Isolation of Nucleotide Sequences Encoding GFP

Peptide sequences from purified GFPs are used to design degenerate oligonucleotide primers for polymerase chain reactions and gene cloning. Protein extracts can be prepared from cnidarian cells by standard methods known to the art. See, for example, Harlow and Lane, *Antibodies, A Laboratory Manual*, Cold Spring Harbor Press, (1988). In a preferred embodiment, cnidarian cells are extracted into a buffer, and the extracts

separated into membrane and soluble fractions. Each fraction is tested for green fluorescence activity in the presence of blue light.

5 Fractions which contain GFP activity are then purified further to determine which components are responsible for the activity. Purification of the active fractions can be carried out by methods known in the art. See, for example, PROTEIN PURIFICATION METHODS - A PRACTICAL APPROACH, Harris et al., Eds. (IRL Press, 10 Oxford, 1989). Extracts prepared as described above are purified by sequential size exclusion chromatography, isoelectric focusing, HPLC size exclusion chromatography, and chromatography on an affinity column. Fractions which display GFP activity can be analyzed further by 15 SDS-PAGE analysis to determine the approximate molecular mass of the active component.

Purified GFPs prepared by the methods described above can be sequenced using methods well known in the art, for example using a gas phase peptide sequencer 20 (Applied Biosystems, Foster City, CA). To determine as much of the peptide sequence as possible, it is preferred that the proteins of the present invention be cleaved into smaller fragments more suitable for gas-phase sequence analysis. This can be achieved by treatment of 25 a purified GFP with selective peptidases, and in a particularly preferred embodiment, with endoproteinase lys-C (Boehringer). The fragments so produced can be separated by reversed-phase HPLC chromatography.

30 The peptide sequences of the proteins determined as above can be used to determine the DNA sequence encoding the protein. Methods for carrying out this determination are well known in the art. See, for example Sambrook et al., MOLECULAR CLONING: A LABORATORY MANUAL, Second Edition, (Cold Spring Harbor Laboratory Press, Cold 35 Spring Harbor, NY 1989).

In a preferred embodiment of the present invention, the peptide sequences are used to design degenerate oligonucleotide primers for polymerase chain reactions.

Each degenerate primer set will preferably contain every possible DNA sequence encoding the corresponding peptide sequences. Primer sets are prepared in both the sense and antisense orientation. Suitable oligonucleotide primers can be synthesized using commercial synthesizers, such as those supplied by Applied Biosystems (Foster City, CA). In a particularly preferred embodiment, the primers include additional nucleotide sequences containing restriction endonuclease cleavage sites. The presence of such sites allows for the directional cloning of PCR products into suitable cloning vectors after treatment with an appropriate restriction enzyme. See Finney, "Molecular Cloning of PCR Products" in CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, Ausubel et al. Eds. (John Wiley & Sons, New York, 1987) p. 15.7.1.

Template DNA for the PCR can be prepared from appropriate cnidarian cells or tissues using methods well known in the art. See Sambrook et al., *supra*. In a preferred embodiment, host cells are crushed under liquid nitrogen and mRNA is extracted using a commercially available kit (Pharmacia, Piscataway, NJ).

The mRNA preparation can then be used as a template for cDNA synthesis using poly(dT) or random hexamer primers by standard techniques. See Sambrook et al., *supra*. In a particularly preferred embodiment, cDNA synthesis is carried out using a commercially available kit (Pharmacia).

The cDNA can then be used directly for PCR using the method of Saiki et al., *Science* 239: 487 (1988). The cDNA also is used to prepare a cDNA library by standard methods. See Sambrook et al., *supra*. In a particularly preferred embodiment, the cDNA is packaged into bacteriophage particles using a commercially available kit (Promega, Madison, WI). The packaged cDNA is then transfected into *E. coli* to produce a cDNA library.

In an alternative preferred embodiment, genomic DNA from cnidarian cells or tissue can be used as the template DNA for the PCR. Genomic DNA can be prepared by

standard methods and PCR can then be used to prepare double stranded DNA molecules to probe the cDNA library and the genomic DNA for the gene(s) encoding GFP. In a preferred embodiment, degenerate primers are prepared corresponding to the termini of the longest peptide sequence determined by peptide sequencing. In a particularly preferred embodiment, primers are used in a PCR with first strand cDNA as template to amplify the DNA encoding the peptide. PCR is carried out under standard conditions. See Sakai et al., *supra*.

PCR amplification products are analyzed by polyacrylamide gel electrophoresis using standard methods. If an amplification product of the expected size (based on the peptide sequence) is found, the product is digested with appropriate restriction enzymes, ligated into a cloning vector and cloned by standard methods. See Sambrook et al, *supra*. In a preferred embodiment, clones are sequenced to verify that sequences according to the expected peptide sequence are present.

Once the DNA sequence encoding the peptide is known, it can be used to prepare non-degenerate primers corresponding to that sequence, again containing restriction enzyme recognition sequences to aid in cloning of DNA products. These primers are used in combination with degenerate primers corresponding to other peptide sequences to generate PCR amplification products which can be cloned and then analyzed as above. By these means, fragments of the gene sequence of the protein can be determined. Alternative methods for carrying out this PCR analysis include use of the 5' or 3' RACE methods using commercially available kits, such as those manufactured by Life Technologies (Gaithersburg, MD) or Clontech (Palo Alto, CA). Primers for this method are selected according to the manufacturer's directions.

Gene fragments prepared as above are excised from the cloning vector by restriction enzyme digestion, labeled with ³²P by conventional methods and used as probes to identify the complete gene encoding the

protein from within a cDNA or genomic library. In a preferred embodiment, the probe is chosen such that it is long enough to ensure hybridization specificity, while remaining short enough to allow reasonable rates of hybridization to the target gene.

A cnidarian genomic DNA library can be prepared by means well-known in the art. See, for example, Slightom et al. "Construction of λ Clone Banks," in METHODS IN PLANT MOLECULAR BIOLOGY AND BIOTECHNOLOGY, Glick et al. (eds.), pages 121-146 (CRC Press, 1993). Genomic DNA can be isolated from cnidarian tissue, for example, by lysing plant tissue with the detergent Sarkosyl, digesting the lysate with proteinase K, clearing insoluble debris from the lysate by centrifugation, precipitating nucleic acid from the lysate using isopropanol, and purifying resuspended DNA on a cesium chloride density gradient. See, for example, Ausubel et al. (eds.), CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, pages 2.3.1-2.3.3 (Wiley Interscience 1990) ["Ausubel"].

DNA fragments that are suitable for the production of a genomic library can be obtained by the random shearing of genomic DNA or by the partial digestion of genomic DNA with restriction endonucleases. See, for example, Ausubel at pages 5.3.2-5.4.4, and Slightom et al., *supra*. Genomic DNA fragments can be inserted into a vector, such as a bacteriophage or cosmid vector, in accordance with conventional techniques, such as the use of restriction enzyme digestion to provide appropriate termini, the use of alkaline phosphatase treatment to avoid undesirable joining of DNA molecules, and ligation with appropriate ligases. Techniques for such manipulation are disclosed by Slightom et al., *supra*, and are well-known in the art. Also see Ausubel at pages 3.0.5-3.17.5.

Screening of the cDNA or genomic library is carried out by conventional methods. See Sambrook et al, *supra*. Clones which hybridize to the probe are purified and their sequences determined. To facilitate sequencing,

5 nested deletions in the clones can be created using standard protocols, or by commercially available kits such as Erase-a-base (Promega, Madison, WI) or The Deletion Factory (Life Technologies, Gaithersburg, MD), following the manufacturer's instructions. The sequences obtained are analyzed for the presence of open reading frames by conventional methods and to check if the entire gene sequence has been found. In a preferred embodiment, cDNA libraries are prepared by both random hexamer and poly (dT) priming from samples, and are used to maximize the chances of finding the complete coding sequence of the desired gene.

10 Once the entire coding sequence of the gene for GFP has been determined, the gene can be inserted into an appropriate expression system. The gene can be expressed in any number of different recombinant DNA expression systems to generate large amounts of protein, which can then be purified.

20 4. Chemical Synthesis of Genes Encoding GFP

Once the amino acid sequence of a GFP is known, a gene encoding this GFP can be synthesized. In addition, the nucleotide sequence of a synthetic gene encoding GFP can be optimized for expression in plants by modifying the codon usage to include plant preferred codons. See, for example, Murray et al., *NAR* 17: 477 (1989). Even more specifically, the nucleotide sequence of a synthetic gene encoding GFP can be optimized for expression in monocotyledonous or dicotyledonous plants. See, for example, Campbell et al., *Plant Physiol.* 92: 1 (1990).

30 Genes encoding GFP can be obtained, for example, by synthesizing the genes with mutually priming long oligonucleotides. See, for example, Ausubel at pages 8.2.8 to 8.2.13. Also, see Wosnick et al., *Gene* 60:115 (1987). Moreover, current techniques using the polymerase chain reaction provide the ability to synthesize genes as large as 1.8 kilobases in length.

Adang et al., *Plant Molec. Biol.* **21**: 1131 (1993); Bambot et al., *PCR Methods and Applications* **2**: 266 (1993).

5. Identification of Functional Fragments of the GFP

DNA clones can be analyzed using a variety of techniques such as restriction analysis, Southern analysis, primer extension analysis, and DNA sequence analysis. Primer extension analysis or S1 nuclease protection analysis, for example, can be used to localize the putative start site of transcription of the cloned gene. Ausubel at pages 4.8.1-4.8.5; Walmsley et al., "Quantitative and Qualitative Analysis of Exogenous Gene Expression by the S1 Nuclease Protection Assay," in *METHODS IN MOLECULAR BIOLOGY, VOL. 7: GENE TRANSFER AND EXPRESSION PROTOCOLS*, Murray (ed.), pages 271-281 (Humana Press Inc. 1991). Functional fragments of the GFP protein are identified by production of green fluorescence in the presence of blue UV light.

The general approach of such functional analysis involves subcloning DNA fragments of a genomic clone, cDNA clone or synthetic gene sequence into an expression vector, introducing the expression vector into a heterologous host, and screening to detect green fluorescence in the presence of UV to blue light. Methods for generating fragments of a cDNA or genomic clone are well-known. Variants of an isolated DNA encoding GFP can be produced by deleting, adding and/or substituting nucleotides for the isolated nucleotides, for example, the nucleotide sequence of SEQ ID NO: 1. Such variants can be obtained, for example, by oligonucleotide-directed mutagenesis, linker-scanning mutagenesis, mutagenesis using the polymerase chain reaction, and the like. See, for example, Ausubel, pages 8.0.3-8.5.9. Also see generally, McPherson (ed.), *DIRECTED MUTAGENESIS: A PRACTICAL APPROACH*, (IRL Press 1991). Thus, the present invention also encompasses DNA molecules comprising nucleotide sequences that have

substantial sequence similarity with SEQ ID NO: 1 and encode a GFP.

6. Methods for Plant Transformation

The GFP-based screening method for plant transformation of the instant application can be used in conjunction with any method of plant transformation and regeneration. Numerous methods for plant transformation have been developed, including biological and physical, plant transformation protocols. See, for example, Miki et al., "Procedures for Introducing Foreign DNA into Plants" in *Methods in Plant Molecular Biology and Biotechnology*, Glick, B.R. and Thompson, J.E. Eds. (CRC Press, Inc., Boca Raton, 1993) pages 67-88. In addition, expression vectors and in vitro culture methods for plant cell or tissue transformation and regeneration of plants are available. See, for example, Gruber et al., "Vectors for Plant Transformation" in *Methods in Plant Molecular Biology and Biotechnology*, Glick, B.R. and Thompson, J.E. Eds. (CRC Press, Inc., Boca Raton, 1993) pages 89-119.

A. Agrobacterium-mediated Transformation

The most widely utilized method for introducing an expression vector into plants is based on the natural transformation system of *Agrobacterium*. See, for example, Horsch et al., *Science* 227:1229 (1985). *A. tumefaciens* and *A. rhizogenes* are plant pathogenic soil bacteria which genetically transform plant cells. The Ti and Ri plasmids of *A. tumefaciens* and *A. rhizogenes*, respectively, carry genes responsible for genetic transformation of the plant. See, for example, Kado, C.I., *Crit. Rev. Plant. Sci.* 10: 1 (1991). Descriptions of *Agrobacterium* vector systems and methods for *Agrobacterium*-mediated gene transfer are provided by Gruber et al., *supra*, Miki et al., *supra*, and Moloney et al., *Plant Cell Reports* 8: 238 (1989).

B. Direct Gene Transfer

Despite the fact the host range for *Agrobacterium*-mediated transformation is broad, some major cereal crop species and gymnosperms have generally been recalcitrant to this mode of gene transfer, even though some success has recently been achieved in rice. Hiei et al., *The Plant Journal* 6: 271-282 (1994). Several methods of plant transformation, collectively referred to as direct gene transfer, have been developed as an alternative to *Agrobacterium*-mediated transformation.

A generally applicable method of plant transformation is microprojectile-mediated transformation wherein DNA is carried on the surface of microprojectiles measuring 1 to 4 μ m. The expression vector is introduced into plant tissues with a biolistic device that accelerates the microprojectiles to speeds of 300 to 600 m/s which is sufficient to penetrate plant cell walls and membranes. Sanford et al., *Part. Sci. Technol.* 5: 27 (1987), Sanford, J.C., *Trends Biotech.* 6: 299 (1988) Sanford, J.C., *Physiol. Plant* 79: 206 (1990), Klein et al., *Biotechnology* 10: 268 (1992).

Another method for physical delivery of DNA to plants is sonication of target cells. Zhang et al., *Bio/Technology* 9: 996 (1991). Alternatively, liposome or spheroplast fusion have been used to introduce expression vectors into plants. Deshayes et al., *EMBO J.*, 4: 2731 (1985), Christou et al., *Proc Natl. Acad. Sci. U.S.A.* 84: 3962 (1987). Direct uptake of DNA into protoplasts using CaCl_2 precipitation, polyvinyl alcohol or poly-L-ornithine have also been reported. Hain et al., *Mol. Gen. Genet.* 199: 161 (1985) and Draper et al., *Plant Cell Physiol.* 23: 451 (1982). Electroporation of protoplasts and whole cells and tissues have also been described. Donn et al., In Abstracts of VIIth International Congress on Plant Cell and Tissue Culture IAPTC, A2-38, p 53 (1990); D'Halluin et al., *Plant Cell* 4: 1495-1505 (1992) and Spencer et al., *Plant Mol. Biol.* 24: 51-61 (1994).

A preferred method is microprojectile-mediated bombardment of immature embryos. The embryos can be bombarded on the embryo axis side to target the meristem at a very early stage of development or bombarded on the scutellar side to target cells that typically form callus and somatic embryos. Targeting of the scutellum using projectile bombardment is well known to those in the art of cereal tissue culture. Klein et al., *Bio/Technol.*, 6: 559-563 (1988); Kartha et al., *Plant Cell Rep.* 8: 429-432 (1989); Sautter et al., *Bio/Technol.*, 9: 1080-1085 (1991); Chibbar et al., *Genome*, 34: 435-460 (1991). The scutellar origin of regenerable callus from cereals is well known. Green et al., *Crop Sci.*, 15: 417-421 (1975); Lu et al., *TAG* 62: 109-112 (1982); and Thomas and Scott, *J. Plant Physiol.* 121: 159-169 (1985). Targeting the scutellum and then using chemical selection to recover transgenic plants is well established in cereals. D/Halluin et al., *Plant Cell* 4: 1495-1505 (1992); Perl et al., *MGG* 235: 279-284 (1992); Cristou et al., *Bio/Technol.* 9: 957-962 (1991). This literature reports DNA targeting of the scutellum and recovery of transgenic callus, plants and progeny based on a chemical selection regime. None of these references teach successful plant transformation wherein transformed cells are visualized with a screenable marker such as GUS.

A preferred transformation method involves bombardment of the scutellar surface of immature embryos to introduce a fGP expression cassette and any other cotransformed genes. Embryos can be pretreated for 1 to 48 hours with a high osmoticum medium or left on a high-osmoticum medium for 24-48 hours after bombardment to improve cell survival and transformation frequencies. Immature embryos are then cultured on typical callus-inducing medium with no selective agent. At each subculture transfer, i.e., every two weeks, the culture is monitored using UV-blue light for GFP fluorescence. Fluorescing calli are separated from non-fluorescing

callus, and grown to the point where plants can be regenerated through standard media progressions.

Many exemplary variations, in terms of target cells and tissue culture routes are known in the art that could be used with GFP in this fashion. Some of these variations include introduction of GFP expression into protoplasts, suspension cells, type II callus, and type I callus, as in Morocz et al., *Theor. Appl. Genet.* 80: 721-726 (1990), Gordon-Kamm et al., *Plant Cell* 2: 603-618 (1990), Fromm et al., *Bio/Technol.* 8: 833-839 (1990) and D'Halluin et al., *Plant Cell* 4: 1495-1505 (1992). Microspores and microspore-derived embryogenic callus also represent a feasible alternative target/culturing route for using GFP screening to recover transformants. See Sun et al., *Plant Cell Rep.* 8: 313-316 (1989) and Mitchell et al., *J. Plant Physiol.* 37: 530-536 (1991). Furthermore, it is expected that the method of the present invention is also be applicable to any dicot species that is known to have a callus stage, for example, tobacco, canola or soybean.

In the case of the maize meristem target, the method entails selective enlargement of transgenic sectors, toward genetic homogeneity, in cell layers that contribute to germline transmission. This method is described in copending U.S. patent application 08/483,091, incorporated herein by reference, and comprises the steps of (A) introducing foreign DNA into a meristem that is not enclosed by sheathing leaves; (B) inducing reorganization of the meristem to increase transgenic sector size, whereby the likelihood that a transgenic sector will contribute to germline transmission is increased; and (C) exposing the meristem to conditions under which it differentiates to form a plantlet, wherein the plantlet contains the transgenic sector or is homogeneously transformed by the foreign DNA, such that the plantlet can be grown into a transformed cereal plant that will transmit the foreign DNA to progeny. The foreign DNA can be introduced into

a plurality of meristems, at least some of which differentiate in step (C) to form a plurality of plantlets.

In one preferred embodiment, reorganization is effected through at least one manipulation selected from the group consisting of (i) imposition of a nonlethal selective pressure on the meristems, (ii) mechanically-induced meristem reorganization, and (iii) hormonally-induced shoot multiplication. In another preferred embodiment the conditions in step (C) are such that the meristems undergo maturation and plant differentiation to form shoot apices, and the method further comprises effecting reorganization of meristem tissue in the shoot apices to enlarge transformed sectors or to produce periclinal L2 chimeras. The reorganization in this regard can be effected, for example, by exposing the shoot apices to nonlethal selection pressure such that transformed cells have a competitive growth advantage over nontransformed cells in the shoot apices, and the proportion of transformed cells in the shoot apices is increased. In yet another preferred embodiment, the method further comprises a step before step (B), e.g., before step (A), of wounding the apical dome selectively. A method of the present invention also can comprise the further steps of (i) dissecting out an axillary bud from above the base of a leaf of a plantlet when a chimeric sector is observed in a substantial portion of the leaf, and then (ii) germinating the axillary bud into a whole plant or subjecting the axillary bud to shoot multiplication.

Likewise, the introduction of GFP-encoding DNA into dicot meristems provides an efficient method of mapping transgenic sectors and following these up into a germline and into progeny. Using mature imbibed seeds as the starting material, the meristem is exposed by removal of the cotyledons, and DNA is introduced into meristem cells. A preferred method for introducing DNA into meristem cells is *Agrobacterium*-mediated delivery, but

other DNA delivery methods can be used with this explant such as bombardment with DNA-coated particles, or silicon carbide fiber-mediated delivery. Kaeppler et al., *Plant Cell Rep.* 9: 415-418 (1990). After DNA introduction, the meristem explants are cultured.

Plants can be manipulated, for example, by removal of the apical meristem, to stimulate axillary or secondary buds which can exhibit larger transgenic sectors relative to the primary shoot. Flowers above transgenic shoots are pollinated and the progeny are analyzed for transgene presence and expression. A variety of starting explants can regenerate shoots in sunflower, and thus represent alternative targets for GFP-encoding DNA delivery and transmission to progeny. These include the seedling meristem (as above), also the seedling hypocotyl, the mature cotyledon, the immature cotyledon, zygotic immature embryos, somatic embryos, and primary leaflets. See for example, respectively, Greco et al., *Plant Sci. Lett.* 36: 73-77 (1984); Krauter et al., *Helia* 14: 117-122 (1991); Power Am. J. Bot. 74: 497-503 (1987); Krauter et al., *Theor. Appl. Genet.* 82: 521-525 (1991); Finer, *Plant Cell Rep.* 6: 372-374 (1987), and Greco et al., *Plant Sci. Lett.* 36: 73-77 (1984).

In similar fashion, GFP expression could be used in other dicot species, for example, soybean, to map sectors and/or identify transformants and follow the transgenes through to progeny. A GFP-encoding cassette and other cotransformed genes can be introduced into soybean cotyledonary node cells using *Agrobacterium*. The explants are cultured on Gamborg (B5) medium with or without kanamycin selection. With no selection, GFP expression is used to identify transgenic chimeric or homogeneously transformed plantlets, which are rooted, grown to maturity and pollinated in the greenhouse. Progeny seed are collected for analysis. The use of cotyledonary nodes and *Agrobacterium* is presented by way of example, but other target cells and culturing methods could be used with GFP for soybean. In addition to the

cotyledonary node cells, mature cotyledons and immature cotyledons have both been used for transformation. See Hinchee et al., *Bio/Technol.* 6: 915-922 (1988) and Parrott et al., *Plant Cell Rep.* 7: 615-617 (1989). Meristems from immature soybean seed embryonic axes have also been used for bombardment-mediated soybean transformation. See Christou et al., *Plant Physiol.* 87: 671-674 (1988) and McCabe et al., *Bio/Technol.* 6: 923-926 (1988).

7. Promoters

A. Inducible Promoters

An inducible promoter is operably linked to a nucleotide sequence encoding a GFP. Optionally, the inducible promoter is operably linked to a nucleotide sequence encoding a signal sequence which is operably linked to a nucleotide sequence encoding GFP. With an inducible promoter the rate of transcription increases in response to an inducing agent.

Any inducible promoter can be used in the instant invention. See Ward et al. *Plant Mol. Biol.* 22: 361-366 (1993). Exemplary inducible promoters include that from the ACE1 system which responds to copper (Mett et al. *PNAS* 90: 4567-4571 (1993)); In2 gene from maize which responds to benzenesulfonamide herbicide safeners (Hershey et al., *Mol. Gen. Genetics* 227: 229-237 (1991) and Gatz et al., *Mol. Gen. Genetics* 243: 32-38 (1994)) or Tet repressor from Tn10 (Gatz et al., *Mol. Gen. Genet.* 227: 229-237 (1991)). A particularly preferred inducible promoter is a promoter that responds to an inducing agent to which plants do not normally respond. An exemplary inducible promoter is the inducible promoter from a steroid hormone gene the transcriptional activity of which is induced by a glucocorticosteroid hormone. Schena et al., *Proc. Natl. Acad. Sci. U.S.A.* 88: 10421 (1991).

The expression vector comprises an inducible promoter operably linked to a nucleotide sequence encoding GFP. The expression vector is introduced into plant cells and presumptively transformed cells are exposed to an inducer of the inducible promoter. The cells are screened for the presence of GFP protein by means of illuminating the cells with UV to blue light and screening for the presence of green fluorescence.

B. Tissue-specific or Tissue Preferred Promoters

A tissue-specific promoter is operably linked to a nucleotide sequence encoding a GFP. Optionally, the tissue-specific promoter is operably linked to a nucleotide sequence encoding a signal sequence which is operably linked to a nucleotide sequence encoding GFP. Plants transformed with a gene encoding GFP operably linked to a tissue-specific promoter produce the GFP protein exclusively, or preferentially, in a specific tissue.

Any tissue-specific or tissue-preferred promoter can be utilized in the instant invention. Exemplary tissue-specific or tissue-preferred promoters include a root-preferred promoter such as that from the phaseolin gene (Murai et al., *Science* **223**: 476-482 (1983) and Sengupta-Gopalan et al., *Proc. Natl. Acad. Sci. USA* **82**: 3320-3324 (1985)); a leaf-specific and light-induced promoter such as that from *cab* or *rubisco* (Simpson et al., *EMBO J.* **4**(11): 2723-2729 (1985) and Timko et al., *Nature* **318**: 579-582 (1985)); an anther-specific promoter such as that from *LAT52* (Twell et al., *Mol. Gen. Genet.* **217**: 240-245 (1989)); a pollen-specific promoter such as that from *Zml3* (Guerrero et al., *Mol. Gen. Genet.* **224**: 161-168 (1993)) or a microspore-preferred promoter such as that from *apg* (Twell et al., *Sex. Plant Reprod.* **6**: 217-224 (1993)).

The expression vector comprises a tissue-specific or tissue-preferred promoter operably linked to a nucleotide sequence encoding GFP. The expression vector is

introduced into plant cells. The cells are screened for the presence of GFP protein by means of illuminating the cells with UV to blue light and screening for the presence of green fluorescence.

C. Constitutive Promoters

A constitutive promoter is operably linked to a nucleotide sequence encoding a GFP or the constitutive promoter is operably linked to a nucleotide sequence encoding a signal sequence which is operably linked to a nucleotide sequence encoding GFP.

Many different constitutive promoters can be utilized in the instant invention. Exemplary constitutive promoters include the promoters from plant viruses such as the 35S promoter from CaMV (Odell et al., *Nature* 313: 810-812 (1985) and the promoters from such genes as rice actin (McElroy et al., *Plant Cell* 2: 163-171 (1990)); ubiquitin (Christensen et al., *Plant Mol. Biol.* 12: 619-632 (1989) and Christensen et al., *Plant Mol. Biol.* 18: 675-689 (1992)); pEMU (Last et al., *Theor. Appl. Genet.* 81: 581-588 (1991)); MAS (Velten et al., *EMBO J.* 3: 2723-2730 (1984)) and maize H3 histone (Lepetit et al., *Mol. Gen. Genet.* 231: 276-285 (1992) and Atanassova et al., *Plant Journal* 2(3): 291-300 (1992)).

The ALS promoter, a XbaI/NcoI fragment 5-prime to the *Brassica napus* ALS3 structural gene (or a nucleotide sequence that has substantial sequence similarity to said XbaI/NcoI fragment), represents a particularly useful constitutive promoter. Co-pending Pioneer Hi-Bred International US Patent Application 08/409,297.

The expression vector comprises a constitutive promoter operably linked to a nucleotide sequence encoding GFP. The expression vector is introduced into plant cells and presumptively transformed cells are screened for the presence of GFP protein by means of illuminating the cells with UV to blue light and screening for the presence of green fluorescence.

8. Signal Sequences For Targeting Proteins to Subcellular Compartments

Transport of GFP to a subcellular compartment such as the chloroplast, vacuole, peroxisome, glyoxysome, cell wall or mitochondrion, or for secretion into the apoplast, is accomplished by means of operably linking the nucleotide sequence encoding a signal sequence to the 5-prime and/or 3-prime region of a gene encoding GFP.

Targeting sequences at the 5-prime and/or 3-prime end of the structural gene may determine, during protein synthesis and processing, where the encoded protein is ultimately compartmentalized. The presence of a signal sequence directs a polypeptide to either an intracellular organelle or subcellular compartment or for secretion to the apoplast.

In addition to the targeting sequences described above, the addition of amino acids to the encoded protein (fusing GFP to either another protein, a protein fragment, or a peptide) can further influence the fate of GFP. For example, a nuclear localization signal (NLS) alone is not sufficient to accumulate GFP in the nucleus. The GFP protein is sufficiently small that it freely diffuses from the nucleus into the cytoplasm, probably through the nuclear pores. The fusion of GFP to a sufficiently large protein permits accumulation in the nucleus if the GFP protein also contains an NLS at the amino-terminus. GFP can also be targeted to the nucleus by fusing GFP to proteins that are normally targeted to the nucleus.

The following exemplary signal sequences were utilized for targeting of GFP. However, the invention encompasses use of any signal sequence, or combination of signal sequences, to facilitate transport of GFP to a subcellular compartment or the apoplast where the protein is not toxic to the cell but can be detected.

More specifically, GFP is expressed in plant cells transformed with a expression vector carrying a

nucleotide sequence encoding a signal sequence(s), which directs a protein to a subcellular compartment or to the apoplast, operably linked to a gene encoding GFP. This GFP gene construct can be expressed from a constitutive, tissue-specific, tissue-preferred or inducible promoter. GFP is transported to a subcellular compartment or the apoplast where it is not toxic to the plant cell. Cells illuminated with UV to blue light can be detected because they emit green fluorescence. The transformed cells are selected for regeneration. If the expression vector also carries a gene encoding a foreign protein or agronomically useful protein, the regenerated plant can be used for production of this protein.

A. PHP8080 - Ubi::CTP-GFPm

A chloroplast signal sequence from the maize cab-m7 has been characterized and the nucleotide sequence is shown below. Becker et al., *Plant Mol. Biol.* **20**: 49 (1992). The signal sequence includes 21 amino acids of the cab-m7 protein that extends beyond the processing site (marked by *).

```

ATG GCG TCC TCC ACG ATG GCC CTC TCC TCC ACC GCC TTC GCC
M  A  S  S  T  H  A  L  S  S  T  A  F  A

GCC AAG GCC GTG AAC GTG CCG TCG TCG TCC GCC TTC GAG
G  K  A  V  N  V  P  S  S  S  A  F  E

GCC CGC GTG ACC ATG AGG AAG ACG GCG GCG AAG GCC AAG
A  R  V  T  *  M  R  X  T  A  A  K  A  K  K

CCA GCT GCG GCG TCC GGG AGC CCG TGG TAC GGC CCC ATG
P  A  A  A  S  G  S  P  W  Y  G  P  M

```

B. PHP8087 - CPN60::GFPm

The mitochondrial signal sequence from the maize chaperonin 60 gene is shown below. Close, P.S., Master's Theses, Iowa State University (1993). The signal sequence includes intron 1 of cpn60II (marked by -) and

includes 2 amino acids from the cpn60II protein beyond the processing site (marked by *).

```

5  ATG TAC CGC GCG GCC GCT AGC CTC GCC TCC AAG GCG CGG CAA GCC GGG AGC
   M Y R A A A S L A S K A R Q A G S
   AGC TCC GCC GCT CGC CAG-gtgaagcagactcgtgttatcacgcgtgtgatgggtctgatg
   S S A A R Q
10 gaggatcccgcccccagatttggtaattgtggcctgggggttcttagtagttctgcgtgcagggtctgtg
   ggttgcgtcatgggtgctgtttattttgtggcgatctgcgggaatctgtagttcgtcgcgcaaaa
   tctaaagctagctcgtcaatggcgtacttggcgtgggttttcacattaatctacgggtggaactcgtc
   actaccgtctctccagttagctgttagacaccgaatacactgattggcagttgagaaacattgatctg
   atccagcagaaatcgatgtcttctggaattcgtttattttatgtcgtgtaacctggggcatggcag
15 tcttcaattgatcaegcactcaccctctgtgtgtgtgatgctttatag-GTT GGA AGC AGG CTT
                                   V G S R L
   GCC TGG AGC AGG AAC TAT GCT GCC ATG
   A W S R N Y *A A M

```

C. PHP8144 - Ubi::BAA-GFPm - Cloned sequence includes 1 amino acid beyond processing site (marked by *, lower case bases show linker sequence).

The barley alpha amylase 1 signal sequence is shown below. Knox, C., et al., "Structure and Organization of Two Divergent Alpha-Amylase Genes From Barley", *Plant Mol. Biol.* 9:3-17 (1987). The cloned sequence includes sequence only up to processing site.

```

35 ATG GCC AAC AAG CAC CTG AGC CTC TCC CTC TTC CTC GTG CTC CTC GCC CTC TCC
   M A N K H L S L S L F L V L L G L S
   GCC TCC CTC GCC TCC GGA tcc ATG
   A S L A S *G S M

```

D. PHP8757 - Ubi::BAA-GFPm-BLVT - Signal sequence starts at *, lower case bases show linker sequence.

The barley lectin vacuole signal sequence is shown below. Lerner et al., *Plant Physiol.* 91: 124-129 (1989).

```

45 AAG atc tac GTG TTC GCC GAG GCC ATC GCC GCC AAC TCC ACC CTC GTG GCC GAG
   K I Y *V F A E A I A A N S T L V A E

```

E. PHP8758 - Ubi::BAA-GFPm-HDEL - Signal sequence starts at *, lower case bases show linker sequence.

The HDEL endoplasmic reticulum retention signal from maize b-70 is shown below. Fontes et al., Plant Cell 3: 483-496 (1991).

```
AAG atc tac GAC GGC GGC GTG GAC GAC GAC CAC GAG CTC
K I Y *D G G V D D D H D E L
```

F. PHP8759 - Ubi::BAA-SVT-GFPm - Marked by ^, lower case bases show linker sequence.

The sweet potato sporamin vacuole signal sequence is shown below. Matsuoka et al., Proc. Natl. Acad. Sci. 88: 834 (1991). The sequence shown includes 4 additional sporamin amino acids beyond the processing site (marked by *).

```
ATG GCC AAC AAG CAC CTG AGC CTC TCC CTC TTC CTC GTG CTC CTC GGC CTC TCC
M A N K H L S L S L F L V L L G L S
GCC TCC CTC GCC TCC GGA CAC AGC AGG TTC AAC CCC ATC AGG CTG CCC ACC ACC
A S L A S G *H S R F N P I R L P T T
CAC GAG CCC GCC AGC AGC GAG ACC gga tcc ATG
H E P A *S S E T G S M
```

G. PHP8882 - Ubi::GFPm-SKL - (marked by *)

The consensus peroxisome signal sequence designated SKL is shown below. Gould et al., J. Cell Biol 108: 1657 (1989).

```
AAG TCG AAG CTT
K *S K L
```

H. Ubi::GR-GFPm

The pea glutathione reductase mitochondrial and chloroplast signal sequence is shown below. Creissen et al., Plant J. 2: 129 (1991). The sequence shown includes 20 additional amino acids beyond the putative processing site (marked by *).

ATG AAC CAG GCG ATG GCC ACC CCG CTG TCC CTG TCC TCC TCC CCG ACC CTG
 M N Q A M A T P L S L S C C S P T L
 ACC AGG TCC ACC CTG TTC TTC ACC AAG ACC TTC CCG TTC TCC CGC TCC TTC TCC
 T R S T L F F T K T F P F S R S P S
 ACC CCG CTG CCG CTG TCC ACC AAG ACC CTG ATC TCC CTG TCC CCG CCG CAC AGG
 T P L P L S T K T L I S L S P P H R
 ACC TTC GCC GTG AGG GCT GAG TCC CAG AAC GGC GCG CAC CCG GCC AGG CAG TAC
 E S Q N G A D P A R Q Y D F D L F T
 GAC TTC GAC CTG TTC ACC ATC GGC
 I G T F A V *R A

I. PHP9053 - Ubi::NLS-GFPm-MALS

The nuclear localization signal from simian virus 40 (SV40) was fused to the N terminus of GFPm. In order to retain the protein in the nucleus the molecular weight of NLS-GFPm was increased by making a carboxy terminal addition of the large unrelated protein maize acetolactate synthase (ALS) (lower case bases show linker sequence). Kalderon, D., Robers, B., Richardson, W., and Smith A., "A short amino acid sequence able to specify nuclear location", *Cell* **39**: 499-509 (1984).

CCG CCC AAG AAG AAG CGC AAG GTG ccc ATG ...
 P P K K K R K V P M
 AAG atc cac ATG
 K I H M

J. Ubi::GFPm-HRGP

A carboxy terminal fusion of a portion of the maize HRGP coding sequence to GFPm was used to anchor GFP to the cell wall. Sequence encoding amino acids 177 to 328 was used. Stiefel, V., Ruiz-Avila, L., Raz R., Valles M., Gomez J., Pages M., Martinez-Izquierdo J., Ludevid M., Landale J., Nelson T., and Puigdomenech P., "Expression of a maize cell wall hydroxyproline-rich glycoprotein gene in early leaf and root vascular differentiation", *Plant Cell* **2**: 785-793 (1990).

TAC ACT CCA AGC CCT AAG CCA CCG GCT ACC AAG CCT CCC AGC
 Y T P S P K P P A T K P P T

CCC AAG CCG ACC CCG CCA ACG TAC ACC CCT TCG CCA AAG CCT
 P K P T P P T Y T P S P K P
 CCG ACA CCC AAG CCG ACC CCG CCG ACG TAC ACC CCT TCT CCC
 P T P K P T P P T Y T P S P
 AAG CCT CCG ACG CCC AAG CCG ACC CCG CCG ACG TAC ACT CCA
 K P P T P K P T P P T Y T P
 AGC CCC AAG CCT CCC ACA CAC CCG ACG CCC AAG CCG ACC CCA
 S P K P P T H P T P K P T P
 CCG ACG TAC ACC CCT TCC CCA AAG CCT CCG ACG CCC AAG CCG
 P T Y T P S P K P P T P K P
 ACC CCA CCG ACG TAC ACC CCT TCC CCA AAG CCT CCG ACA CCC
 T P P T Y T P S P K P P T P
 AAG CCG ACC CCA CCG ACG TAC ACC CCT TCC CCA AAG CCT CCG
 K P T P P T Y T P S P K P P
 ACA CCC AAG CCG ACC CCA CCG ACG TAC ACT CCC ACA CCG AAG
 T P K P T P P T Y T P T P K
 CCG CCG GCC ACC AAG CCG CCC ACC TAC ACT CCG ACG CCG CCG
 P P A T K P P T Y T P T P P
 GTG TCT CAC ACC CCC AGC CCG CCG CCA CCA TAC TAC
 V S H T P S P P P P Y Y

K. Ubi::GFPm-SOD

Superoxide dismutase coding sequence fused to
 GFPm. Alcendor D., Chapman G., and Beaman B.,
 "Isolation, sequencing and expression of the
 superoxide dismutase-encoding gene (sod) of *Nocardia*
asteroides strain GUH-2", *Gene* 164: 143-147 (1995).

GTG GCT GAG TAC ACG CTG CCG GAT CTG GAT TAC GAC TAC AGC
 V A E Y T L P D L D Y D Y S
 GCC TGG AAC CCC ACA TCT CCG GGC AGA TCA ACG AGC TGA CAC
 A L E P H I S G Q I N E L H
 CAT TCC AAG CAC CAC GCC GCC TAC GTC GCC GGT GCC AAC ACG
 H S K H H A A Y V A G A N T
 GCA CTG GAG AAG CTG GAA GCC GCC CGT GAG GCC GGC GAT CAC
 A L E K L E A A R E A G D H
 AGC GCG ATC TTC CTG CAC GAG AAG AAC CTC GCG TTC CAC CTC
 S A I F L H E K N L A F H L
 GGC GGA CAC GTC AAC CAC TCC ATC TGG TGG AAG AAC CTG TCC
 G G H V N H S I W W K N L S
 CCC AAC GGT GGC GAC AAG CCG GTC GGC GAG CTG GCC GCG GCC
 P N G G D K P V G E L A A A
 ATC GAC GAC CAG TTC GGT TCG TTC GAC AAG TTC GCG GCG CAG
 I D D Q F G S F D K F R A Q
 TTC ACC GCC GCG GCC AAC GGC CTG CAG GGC TCG GGC TGG GCG
 F T A A A N G L Q G S G W A
 GTG CTC GGT TAC GAC ACC CTC GGC CAG AAG CTG CTG ACC TTC
 V L G Y D T L G Q K L L T F
 CAG CTC TAC GAC CAG CAG GCC AAC GTG CCG CTG GGC ATC ATC
 Q L Y D Q Q A N V P L G I I
 CCG CTG CTC CAG GTC GAC ATG TGG GAG CAC GCC TTC TAC CTG
 P L L Q V D M W E H A F Y L
 CAG TAC AAG AAC GTC AAG GCC GAC TAC GTG ACC GCG TTC TGG
 Q Y K N V K A D Y V T A F W

AAC GTC GTC AAC TGG GCC GAC GTG CAG GAC CGC TTC GGC AAG
N V V N W A D V Q D R F G K
GCC GTC AAC CAG GCC AAG GGC CTT ATC TTC GGG
A V N Q G K G L I F G

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9. Detection of GFP in Transformed Plant Cells

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GFP is detected in transformed plant cells by conventional spectrophometric methods. The transformed cells or tissue are screened for the presence of GFP protein by means of the illuminating the cells with UV to blue light and screening for the presence of green fluorescence.

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Compound and dissecting microscopes were fitted with appropriate filter combinations for fluorescent protein excitation. Illumination with UV-blue light (exciting around the absorption maximum of 395 nm or around the minor peak at approximately 475 nm for GFP; around 480-490 nm for the red-shifted version and around 380 nm for the blue fluorescent protein) is required for visualization. A hand-held lamp for benchtop work also permits good visualization. Cut-off filters or bandpass filters between the fluorescing tissue and the viewer (i.e. around the intermediate objective or the eyepieces of the microscope, or hand-held in front of the eyes if working on the benchtop) greatly reduced background autofluorescence from the tissue. For GFP and the red-shifted GFP this cut-off or bandpass filter permitted light between 500-550 nm to reach the viewer. For the blue fluorescent protein, this filter permitted the passage of 420 to 450 nm light. Useful filters and wavelengths are not restricted to those described above, and further generic description of the optical characteristics of this system are available. Heim et al., *Currently Biology* 6: 178-182 (1996).

10. Foreign Protein Genes and Agronomic Genes

With transgenic plants according to the present invention, a foreign protein can be produced in commercial quantities. Thus, the selection and propagation techniques described above yield a plurality of transgenic plants which are harvested in a conventional manner, and a foreign protein then is extracted from a tissue of interest or from total biomass. Protein extraction from plant biomass can be accomplished by known methods which are discussed, for example, by Heney and Orr, *Anal. Biochem.* 114: 92-6 (1981).

According to a preferred embodiment, the transgenic plant provided for commercial production of foreign protein is maize. In another preferred embodiment, the biomass of interest is seed. For the relatively small number of transgenic plants that show higher levels of expression, a genetic map can be generated, primarily via conventional RFLP and PCR analysis, which identifies the approximate chromosomal location of the integrated DNA molecule. For exemplary methodologies in this regard, see Glick and Thompson, *METHODS IN PLANT MOLECULAR BIOLOGY AND BIOTECHNOLOGY* 269-284 (CRC Press, 1993). Map information concerning chromosomal location is useful for proprietary protection of a subject transgenic plant. If unauthorized propagation is undertaken and crosses made with other germplasm, the map of the integration region can be compared to similar maps for suspect plants, to determine if the latter have a common parentage with the subject plant. Map comparisons would involve hybridizations, RFLP, PCR and sequencing, all of which are conventional techniques.

Likewise, by means of the present invention, agronomic genes can be expressed in transformed plants. More particularly, plants can be genetically engineered to express various phenotypes of agronomic interest. The

genes implicated in this regard include, but are not limited to, those categorized below.

1. Genes That Confer Resistance To Pests or Disease And That Encode:

(A) Plant disease resistance genes. Plant defenses are often activated by specific interaction between the product of a disease resistance gene (R) in the plant and the product of a corresponding avirulence (Avr) gene in the pathogen. A plant variety can be transformed with cloned resistance gene to engineer plants that are resistant to specific pathogen strains. See, for example Jones et al., *Science* 266: 789 (1994) (cloning of the tomato Cf-9 gene for resistance to *Cladosporium fulvum*); Martin et al., *Science* 262: 1432 (1993) (tomato Pto gene for resistance to *Pseudomonas syringae* pv. tomato encodes a protein kinase); Mindrinos et al., *Cell* 78: 1089 (1994) (*Arabidopsis* RSP2 gene for resistance to *Pseudomonas syringae*).

(B) A *Bacillus thuringiensis* protein, a derivative thereof or a synthetic polypeptide modeled thereon. See, for example, Geiser et al., *Gene* 48: 109 (1986), who disclose the cloning and nucleotide sequence of a Bt δ -endotoxin gene. Moreover, DNA molecules encoding δ -endotoxin genes can be purchased from American Type Culture Collection (Rockville, MD), under ATCC accession Nos. 40098, 67136, 31995 and 31998.

(C) A lectin. See, for example, the disclosure by Van Damme et al., *Plant Molec. Biol.* 24: 825 (1994), who disclose the nucleotide sequences of several *Clivia miniata* mannose-binding lectin genes.

(D) A vitamin-binding protein such as avidin. See U.S. patent application serial No. 07/911,864, the contents of which are hereby incorporated by reference.

The application teaches the use of avidin and avidin homologues as larvicides against insect pests.

(E) An enzyme inhibitor, for example, a protease inhibitor or an amylase inhibitor. See, for example, Abe et al., *J. Biol. Chem.* 262: 16793 (1987) (nucleotide sequence of rice cysteine proteinase inhibitor), Huub et al., *Plant Molec. Biol.* 21: 985 (1993) (nucleotide sequence of cDNA encoding tobacco proteinase inhibitor I), and Sumitani et al., *Biosci. Biotech. Biochem.* 57: 1243 (1993) (nucleotide sequence of *Streptomyces nitrosporeus* α -amylase inhibitor).

(F) An insect-specific hormone or pheromone such as an ecdysteroid and juvenile hormone, a variant thereof, a mimetic based thereon, or an antagonist or agonist thereof. See, for example, the disclosure by Hammock et al., *Nature* 344: 458 (1990), of baculovirus expression of cloned juvenile hormone esterase, an inactivator of juvenile hormone.

(G) An insect-specific peptide or neuropeptide which, upon expression, disrupts the physiology of the affected pest. For example, see the disclosures of Regan, *J. Biol. Chem.* 269: 9 (1994) (expression cloning yields DNA coding for insect diuretic hormone receptor), and Pratt et al., *Biochem. Biophys. Res. Comm.* 163: 1243 (1989) (an allostatin is identified in *Diploptera punctata*). See also U.S. patent No. 5,266,317 to Tomalski et al., who disclose genes encoding insect-specific, paralytic neurotoxins.

(H) An insect-specific venom produced in nature by a snake, a wasp, etc. For example, see Pang et al., *Gene* 116: 165 (1992), for disclosure of heterologous expression in plants of a gene coding for a scorpion insectotoxic peptide.

(I) An enzyme responsible for an hyperaccumulation of a monterpene, a sesquiterpene, a steroid, hydroxamic acid, a phenylpropanoid derivative or another non-protein molecule with insecticidal activity.

5 (J) An enzyme involved in the modification, including the post-translational modification, of a biologically active molecule; for example, a glycolytic enzyme, a proteolytic enzyme, a lipolytic enzyme, a
10 nuclease, a cyclase, a transaminase, an esterase, a hydrolase, a phosphatase, a kinase, a phosphorylase, a polymerase, an elastase, a chitinase and a glucanase, whether natural or synthetic. See PCT application WO 93/02197 in the name of Scott et al., which discloses the
15 nucleotide sequence of a callase gene. DNA molecules which contain chitinase-encoding sequences can be obtained, for example, from the ATCC under accession Nos. 39637 and 67152. See also Kramer et al., *Insect Biochem. Molec. Biol.* 23: 691 (1993), who teach the nucleotide
20 sequence of a cDNA encoding tobacco hookworm chitinase, and Kawalleck et al., *Plant Molec. Biol.* 21: 673 (1993), who provide the nucleotide sequence of the parsley ubi4-2 polyubiquitin gene.

25 (K) A molecule that stimulates signal transduction. For example, see the disclosure by Botella et al., *Plant Molec. Biol.* 24: 757 (1994), of nucleotide sequences for mung bean calmodulin cDNA clones, and Griess et al., *Plant Physiol.* 104: 1467 (1994), who
30 provide the nucleotide sequence of a maize calmodulin cDNA clone.

35 (L) A hydrophobic moment peptide. See U.S. patent applications serial No. 08/168,809 (disclosure of peptide derivatives of Tachyplesin which inhibit fungal plant pathogens) and serial No. 08/179,632 (teaches synthetic antimicrobial peptides that confer disease resistance),

the respective contents of which are hereby incorporated by reference.

5 (M) A membrane permease, a channel former or a channel blocker. For example, see the disclosure by Jaynes et al., *Plant Sci.* 89: 43 (1993), of heterologous expression of a cecropin- β lytic peptide analog to render transgenic tobacco plants resistant to *Pseudomonas solanacearum*.

10 (N) A viral-invasive protein or a complex toxin derived therefrom. For example, the accumulation of viral coat proteins in transformed plant cells imparts resistance to viral infection and/or disease development
15 effected by the virus from which the coat protein gene is derived, as well as by related viruses. See Beachy et al., *Ann. Rev. Phytopathol.* 28: 451 (1990). Coat protein-mediated resistance has been conferred upon transformed plants against alfalfa mosaic virus, cucumber
20 mosaic virus, tobacco streak virus, potato virus X, potato virus Y, tobacco etch virus, tobacco rattle virus and tobacco mosaic virus. *Id.*

25 (O) An insect-specific antibody or an immunotoxin derived therefrom. Thus, an antibody targeted to a critical metabolic function in the insect gut would inactivate an affected enzyme, killing the insect. Cf. Taylor et al., Abstract #497, SEVENTH INT'L SYMPOSIUM ON MOLECULAR PLANT-MICROBE INTERACTIONS (1994) (enzymatic
30 inactivation in transgenic tobacco via production of single-chain antibody fragments).

35 (P) A virus-specific antibody. See, for example, Tavladoraki et al., *Nature* 366: 469 (1993), who show that transgenic plants expressing recombinant antibody genes are protected from virus attack.

(Q) A developmental-arrestive protein produced in nature by a pathogen or a parasite. Thus, fungal endo α -1,4-D-polygalacturonases facilitate fungal colonization and plant nutrient release by solubilizing plant cell wall homo- α -1,4-D-galacturonase. See Lamb et al., *Bio/Technology* 10: 1436 (1992). The cloning and characterization of a gene which encodes a bean endopolygalacturonase-inhibiting protein is described by Toubart et al., *Plant J.* 2: 367 (1992).

(R) A developmental-arrestive protein produced in nature by a plant. For example, Logemann et al., *Bio/Technology* 10: 305 (1992), have shown that transgenic plants expressing the barley ribosome-inactivating gene have an increased resistance to fungal disease.

2. Genes That Confer Resistance To A Herbicide, For Example:

(A) A herbicide that inhibits the growing point or meristem, such as an imidazalinone or a sulfonylurea. Exemplary genes in this category code for mutant ALS and AHAS enzyme as described, for example, by Lee et al., *EMBO J.* 7: 1241 (1988), and Miki et al., *Theor. Appl. Genet.* 80: 449 (1990), respectively.

(B) Glyphosate (resistance imparted by mutant EPSP synthase and aroA genes, respectively) and other phosphono compounds such as glufosinate (PAT and bar genes), and pyridinoxy or phenoxy proprionic acids and cyclohexones (ACCase inhibitor-encoding genes). See, for example, U.S. patent No. 4,940,835 to Shah et al., which discloses the nucleotide sequence of a form of EPSP which can confer glyphosate resistance. A DNA molecule encoding a mutant aroA gene can be obtained under ATCC accession No. 39256, and the nucleotide sequence of the mutant gene is disclosed in U.S. patent No. 4,769,061 to Comai. European patent application No. 0 333 033 to

Kumada et al. and U.S. patent No. 4,975,374 to Goodman et al. disclose nucleotide sequences of glutamine synthetase genes which confer resistance to herbicides such as L-phosphinothricin. The nucleotide sequence of a phosphinothricin-acetyl-transferase gene is provided in European application No. 0 242 246 to Leemans et al. De Greef et al., *Bio/Technology* 7: 61 (1989), describe the production of transgenic plants that express chimeric bar genes coding for phosphinothricin acetyl transferase activity. Exemplary of genes conferring resistance to phenoxy proprionic acids and cyclohexones, such as sethoxydim and haloxyfop, are the Acc1-S1, Acc1-S2 and Acc1-S3 genes described by Marshall et al., *Theor. Appl. Genet.* 83: 435 (1992).

(C) A herbicide that inhibits photosynthesis, such as a triazine (*psbA* and *gs+* genes) and a benzonitrile (nitrilase gene). Przibilla et al., *Plant Cell* 3: 169 (1991), describe the transformation of *Chlamydomonas* with plasmids encoding mutant *psbA* genes. Nucleotide sequences for nitrilase genes are disclosed in U.S. patent No. 4,810,648 to Stalker, and DNA molecules containing these genes are available under ATCC accession Nos. 53435, 67441 and 67442. Cloning and expression of DNA coding for a glutathione S-transferase is described by Hayes et al., *Biochem. J.* 285: 173 (1992).

3. Genes That Confer Or Contribute To A Value-Added Trait, Such As:

(A) Modified fatty acid metabolism, for example, by transforming a plant with an antisense gene of stearoyl-ACP desaturase to increase stearic acid content of the plant. See Knultzon et al., *Proc. Nat'l Acad Sci. USA* 89: 2624 (1992).

(B) Decreased phytate content

(1) Introduction of a phytase-encoding gene would enhance breakdown of phytate, adding more free phosphate to the transformed plant. For example, see Van Hartingsveldt et al., *Gene* 127: 87 (1993), for a disclosure of the nucleotide sequence of an *Aspergillus niger* phytase gene.

(2) A gene could be introduced that reduces phytate content. In maize, this, for example, could be accomplished, by cloning and then re-introducing DNA associated with the single allele which is responsible for maize mutants characterized by low levels of phytic acid. See Raboy et al., *Maydica* 35: 383 (1990).

(C) Modified carbohydrate composition effected, for example, by transforming plants with a gene coding for an enzyme that alters the branching pattern of starch. See Shiroza et al., *J. Bacteriol.* 170: 810 (1988) (nucleotide sequence of *Streptococcus mutans* fructosyltransferase gene), Steinmetz et al., *Mol. Gen. Genet.* 200: 220 (1985) (nucleotide sequence of *Bacillus subtilis* levansucrase gene), Pen et al., *Bio/Technology* 10: 292 (1992) (production of transgenic plants that express *Bacillus licheniformis* α -amylase), Elliot et al., *Plant Molec. Biol.* 21: 515 (1993) (nucleotide sequences of tomato invertase genes), Søgaard et al., *J. Biol. Chem.* 268: 22480 (1993) (site-directed mutagenesis of barley amylase gene), and Fisher et al., *Plant Physiol.* 102: 1045 (1993) (maize endosperm starch branching enzyme II).

Example 1
Construction of Genes and Vectors

5 The GFPm nucleotide sequence was derived from
a back translation of the protein sequence using maize
preferred codons. Sequence analysis was performed using
the Wisconsin Sequence Analysis Package from Genetics
Computer Group, Madison, WI. The nucleotide sequence was
10 assembled from a series of synthetic oligonucleotides.
Cloning sites include a 5-prime flanking *Bam*HI
restriction site, an *Afl*III site at the start codon, a 3-
prime flanking *Hpa*I site or a *Bgl*II site converting the
stop codon to an isoleucine (Fig. 1).

15 Amino terminal fusions to GFPm were created using
synthetic oligonucleotides encoding the signal sequence
flanked by a 5-prime *Bam*HI or *Bgl*II restriction site and
3-prime *Nco*I site. The synthetic signal sequences were
cloned into PHP7921 (Fig. 4) using the 5-prime flanking
20 *Bam*HI and *Afl*III restriction sites of GFPm to create in-
frame fusions. Sequences cloned in this manner include
the maize chloroplast signal sequence, barley alpha-
amylase signal sequence, pea glutathione reductase signal
sequence, and SV40 nuclear localization signal. The
25 sporamin vacuole signal sequence was synthesized with a
5-prime *Bsp*EI and 3-prime *Bam*HI for insertion between the
BAA pre-sequence and GFPm coding sequence of PHP8144
(Fig. 5). The CPN60 promoter site, created by site-
directed mutagenesis, was used with the GFPm *Afl*III site
30 to make the fusion sequence. Each of plasmids PHP7921
and PHP8144 are based on pUC18.

35 Carboxy terminal sequences were synthesized with
a 5-prime *Bgl*II and 3-prime *Hpa*I site, and cloned into
PHP7921 or PHP8144 using the 3-prime *Bgl*II and *Hpa*I sites
of GFPm. The barley lectin vacuole target, HDEL ER
retention signal, SKL peroxisome signal, ALS coding
sequence and HRGP coding sequence fusions were made using
this strategy.

TABLE 1
PLASMIDS

PHP Number	Target Crop	Promoter	Targeting				Other 5' Sequences	Structural Gene	3' Sequence	Comments
			5' Sequence	3' Sequence	Localized To:					
7956	maize	4xERE	--	--	Cytoplasm		Omega Prime (O') Adhl intron I	GFPm	pinII	Estrogen-inducible expression
7667	maize	ubiquitin	--	--	Cytoplasm		--	GFP(native)	nos	Jellyfish structural gene
7921	maize	ubiquitin	--	--	Cytoplasm		--	GFPm	pinII	Cytoplasmic expression
8489	maize	ubiquitin	--	--	Cytoplasm		--	GFPm	pinII	MAR flanking cassette
8490	maize	ubiquitin	--	--	Cytoplasm		--	GFPm-S65T	pinII	MAR flanking cassette, red-shift
8080	maize	ubiquitin	CTP	--	chloroplast		--	GFPm	pinII	Chloroplast signal sequence, cab-m7 from maize

8087	maize	CPN60	CPN60	--	mitochondria a	--	GFPm	pinII	Targeted to mitochondria, signal from maize chaparonin 60
8144	maize	ubiquitin	BAA	--	secreted	--	GFPm	pinII	Secreted GFP; signal from barley alpha amylase 1
8757	maize	ubiquitin	BAA	BLVT	vacuole	--	GFPm	pinII	Barley lectin vacuole signal seq. on carboxy end of BAA- GFPm
8758	maize	ubiquitin	BAA	HDEL	endoplasmic reticulum	--	GFPm	pinII	HDEL endoplasmic reticulum signal from maize b-70
8759	maize	ubiquitin	BAA	--	vacuole	SVT	GFPm	pinII	Sweet potato sporamin vacuole signal sequence

8882	maize	ubiquitin	--	SKL	peroxisome	--	GFPm	pinII	Consensus peroxisomal signal sequence at carboxy terminus
8986	maize	ubiquitin	T7 TAG-NLS-	--	nucleus	--	GFPm	pinII	Nuclear localization sequence
9053	maize	ubiquitin	T7 TAG-NLS-	--	nucleus	--	MALS-GFPm	pinII	Nuclear localization sequence; protein fusion to prevent escape
9057	maize	ubiquitin	--	--	nucleus	--	MALS-GFPm	pinII	Fusion protein to test effect on GFP as reporter
9058	maize	ubiquitin	--	--	cytoplasm	--	PATm-GFPm	pinII	PAT-GFP fusion to test efficacy

8744	maize	ubiquitin	--	--	nucleus	--	GFPm-ZmRAD51	pinII	Protein fusion: for studies on Rad51 expression/function
8081	maize	ubiquitin	--	--	cytoplasm	FRT	GFPm	pinII	To verify that FRT sequence does not impair expression
8674	maize	2x358	--	--	cytoplasm	O' Adh1 intron	FRT-bar-FRT-GFPm	pinII	Select on bialaphos, then excise bar and activate GFP
8911	maize	ubiquitin / 2x358	--	adh1 intron 1	cytoplasm	Ds	GFPm bat	pinII pinII	ubi-Ds-2x358: bar::pin-Ds-GFPm::pinII
8088	maize	ubiquitin	--	--	cytoplasm	--	GFPm-S65T	pinII	Red-shifted GFPm
8138	maize	ubiquitin	--	--	cytoplasm	--	GFPm-Red2	pinII	Red-shifted version #2 F64M, S65G, Q69L

8175	maize	ubiquitin	--	--	cytoplasm	--	Blue FP	pinII	GFPm-modified Y66H
8007	maize	2x35s ubiquitin					adh1 intron1::bar ubi intron::FLP	pinII pinII	Tandem vector for FLP excision of FRT to activate GFPm
8011	sunflo wer	2x35s ALS	-- --	-- --	cytoplasm	O'	GFPm NPT-II	pinII pinII	Binary
8491	sunflo wer	2x35s ALS	-- --	-- --	cytoplasm	O'	GFPm-S65T NPT-II	pinII pinII	Binary with red-shifted GFP

The ubiquitin promoter is described in Christensen et al., *Plant Mol. Biol.* 18: 675-689 (1992). The 4xERE promoter is described in Klein-Hitpass et al., *Cell* 46: 1053-1061 (1986). The CPN60; 2x35S and ALS promoters are described in Close P. Ph.D. Dissertation, Iowa State University; Gardner et al., *Nucl. Acid Res.* 9: 2871-2888, (1981) and PHI Patent Application 08/409,297, respectively. The Omega prime (O') 5-prime sequence is described by Gallie et al., *Nucl. Acids Res.* 15: 3257-3273 (1987) while the Adh 1 intron I is described by Dennis et al., *Nucl. Acids Res.* 12: 3983-3990 (1984). The 3-prime sequence pinII is described in An et al., *Plant Cell* 1: 115-122 (1989). Structural genes used in the construction of the vectors include the GFP gene (native jellyfish) described by Prasher et al., *Gene* 111: 229-233 (1992); the maize ALS gene described by Fromm et al., *Bio/Technol.* 8: 883 (1990) and the maize HPRG described by Steifel et al., *Plant Mol. Biol.* 11: 483-493 (1988). The FLP/FRT system contained in plasmids PHP8674 and PHP8007 is described in O'Gorman et al., *Science* 251: 1351-1355 (1991) and Jayaram M., *Proc. Natl. Acad. Sci. USA* 82: 5875-5879 (1985). The Ac/Ds system is described by Muller-Neumann et al., *Mol. Gen. Genet.* 198: 19-24 (1984).

For *Agrobacterium*-mediated transformation of sunflower, the backbone of PHP8011 is a bin 19-based vector. Beven, *Nucl. Acids Res.* 12: 8711-8721 (1984). The NPT-II cassette was removed from bin 19 and the following cloned between the left and right border sequences; the double 35S promoter, omega', the GFPm coding sequence and the pinII 3' region (all described above) were cloned downstream of the left border sequence, followed in a head-to-tail orientation by the BnALS3 promoter, the NPT-II coding sequence and the pinII 3' region. Beven, "Binary *Agrobacterium* Vectors for Plant Transformation", *Nucl. Acids Res.* 12: 8711-8721 (1984).

GFP_r, BFP and GFPs (soluble GFP) were constructed by means of oligonucleotide-directed mutagenesis of the GFP_m nucleotide sequence shown in Figure 1 as described by Ausubel, *supra*. With regard to GFP_r, TCC was changed to ACG resulting in the S65T substitution in the GFP amino acid sequence. With regard to BFP, TAC was changed to CAC, resulting in the Y66H substitution in the GFP amino acid sequence.

With regard to GFPs, the following sequence changes in GFP_m were made. TCC was changed to AGC, resulting in a F99S substitution in the GFP amino acid sequence. The oligonucleotide used to introduce this mutation also caused a silent mutation at R96 (AGC changed to CGG), thereby adding a BsrBI restriction site. ATG was changed to ACC, resulting in an M153T substitution in the GFP amino acid sequence. GTG was changed to GCC, resulting in a V163A substitution in the GFP amino acid sequence. A single oligonucleotide was used to change the amino acids at 153 and 163. This oligonucleotide also caused a silent mutation at A154 (GCC changed to GCG), which added a SacII restriction site. As reported by Cramer et al., *supra*, these three amino acid changes did not alter the absorption/emission characteristics of GFP. These three amino acid substitutions, however, increased the solubility of GFP. Maize cells transformed with the GFPs gene fluoresce in blue to UV light based on transient assays.

Example 2**5 Maize Transformation with GFP Screening or Bialaphos Selection**

10 Maize Hi-II immature embryos were co-transformed via microprojectile bombardment with a mixture of PHP7814 (ubi::PAT::pinII) and PHP8409. Plasmid PHP7814 carries the PAT gene encoding resistance to bialaphos operably linked to the ubiquitin promoter and the terminator from the pinII gene. Standard
15 CaCl₂/spermidine precipitation of DNA onto tungsten microprojectiles was used to prepare the DNA-particles. Prior to bombardment, the embryos of approximately 1.5-2.0 mm in length were cultured on 560P medium including N6 salts, Eriksson's vitamins, 0.69 mg/L proline, 2
20 mg/L 2,4-D, and 3% sucrose. After 4-5 days of incubation in the dark at 28°C, embryos were removed from 560P medium and cultured, coleorhizal end up, on 560L medium which is equivalent to 560P except that it contains 12% sucrose. Embryos were allowed to
25 acclimate to this medium for 3 h prior to transformation.

Embryos were transformed using the PDS-1000 Helium Gun from Bio-Rad at one shot per sample using 650 PSI rupture disks. On average approximately 0.0067
30 µg of DNA was delivered per shot. For each treatment (pat gene with bialaphos selection or GFP gene with screening for GFP), 447 embryos were used. For the bialaphos-selection treatment; following bombardment, the embryos were maintained on 560L medium for 48 h
35 before transfer to 560R selection medium containing N6 salts, Eriksson's vitamins, 2 mg/L 2,4-D, 3% sucrose and 3mg/L bialaphos. Plates were maintained at 28°C in the dark and were observed for colony recovery. Transfer of colonies to fresh medium occurred every two
40 weeks. Transgenic colony recovery under bialaphos

selection was accomplished by picking the healthy, growing transgenic calli from amongst the inhibited background of non-transgenic tissue as the material was progressively transferred to fresh media. For the GFP screening treatment, 560P media was used after bombardment and the two-day recovery on 560L. Calli were transferred to fresh non-selective 560P medium every two weeks. Screening for GFP expression was carried out at each transfer using a Xenon and/or Mercury light source with the appropriate filters for GFP visualization.

Table 2

Recovery of Transgenic Events in Maize Hi-II Callus Using Either GFP Visualization or Bialaphos Selection

Selection Method	Number of Events Recovered	
	Initial Events	Events Capable of Regeneration
Bialaphos selection	16	12
Screened using GFP	9	8

Transgenic events were recovered from each treatment. The largest number of transgenic events were consistently obtained by means of bialaphos selection. However, the frequency of recovery of transgenic events with GFP screening was approximately one-half the rate of recovery with bialaphos selection which was surprisingly high considering that GFP screening does not rely on selection of transgenic events in the presence of a cellular toxin.

Once GFP expressing colonies were identified they were monitored regularly for new growth and expression using the Xenon light source. Plant cells containing GFP were regenerated by transferring the callus to 288 medium containing MS salts, 1 mg/L IAA,

0.5 mg/L zeatin and 4% sucrose. The callus was placed in the light. As plantlets developed they were transferred to tubes containing 272K, hormone-free MS medium and 3% sucrose. The percentage of green
5 fluorescent colonies that regenerated into whole plants was determined.

Leaf samples were collected from TO transgenic plants from events recovered by means of bialaphos selection, or for events recovered by means of GFP
10 visualization (non-selected). Genomic DNA was extracted from all leaf samples and Southern analysis was performed using the PAT and GFP genes as hybridization probes. Hybridization for the PAT gene and GFP gene were performed sequentially on the same
15 blots, by stripping and reprobing for the other transgene. Bands hybridizing for the PAT gene and for the GFP gene were observed in all 8 GFP-screened events tested and in all 12 of the bialaphos selected events tested. Copy numbers and integration patterns for both
20 treatments appeared similar, and were consistent with previous experiments by the inventors using bialaphos selection of callus. Copy numbers ranged from low (single or two copies for each gene) to numerous (i.e., >5 copies per gene). Integration events were
25 predominantly at single integration sites but in some cases integration at multiple locations in the genome was observed.

Bialaphos is the most efficient chemical selective agent for maize. Consequently, the high
30 frequency with which transgenic events were obtained with GFP screening, compared to bialaphos selection, was unexpected. An advantage of GFP screening over bialaphos selection is that non-transformed tissue is not killed. Accordingly, transgenic sectors containing
35 the GFP gene develop in a healthier environment for growth and development.

Example 3**The GFP Marker in Maize Cytoplasm, Chloroplast and Mitochondria**

5 Maize A188 x B73-derived cells were transformed
with plasmids PHP8080, PHP7921 and PHP8087. Each of
these plasmids is described in Example 1 and Table 1.
Plasmid PHP8080 carries the GFPm gene operably linked
10 to the ubiquitin promoter and a chloroplast signal
sequence. GFP is directed to the chloroplasts of cells
transformed with PHP8080. Plasmid PHP7921 carries the
GFPm gene operably linked to ubiquitin promoter. GFP
remains in the cytoplasm of cells transformed with
15 pPGP7921. Plasmid PHP8087 carries the GFPm gene
operably lined to the CPN60 promoter and CPN60 signal
sequence. GFP is directed to mitochondria of cells
transformed with pPGP8087.

 The cells were transformed by culturing maize
embryos approximately 1.5 to 2 mm in length onto 560P
20 medium containing N6 salts, Eriksson's vitamins, 0.69
g/L Proline, 2 mg/L 2,4-D and 3% sucrose. After 4-5
days of incubation in the dark at 28°C, embryos were
removed from 560P medium and cultured, coleorhizal end
up, onto 560L medium which is equivalent to 560P but
25 contains 12% sucrose. Embryos were allowed to
acclimate to this medium for 3 h prior to
transformation. Embryos were transformed using the
PDS-1000 Helium Gun from Bio-Rad at one shot per sample
using 650PSI rupture disks. DNA delivered per shot
30 averaged at 0.0667 µg. Following bombardment, all
embryos were maintained on 560L medium for 48 hours
before transfer to 3% sucrose and 3 mg/L bialaphos.
Plates were maintained at 28°C in the dark and were
observed for colony recovery with transfers to fresh
35 medium occurring every two weeks. Transgenic colony
recovery was noted initially as growing callus tissue
with a healthy phenotype on selection. Screening for

GFP expression was completed with each transfer using a Xenon light source with UV filter attachments.

GFP localization was confirmed by analysis of TO callus. Samples of calli from the targeted GFP transformations were fixed in FAA and then examined using an inverted microscopes with UV filters to visualize GFP. For each targeted construct GFP expression was localized to the specified organelle.

Once GFP expressing colonies were identified they were monitored regularly for new growth and expression using the Xenon light source. Plant cells containing GFP were regenerated by transferring the callus to 288 medium containing MS salts, 1 mg/L IAA, 0.5 mg/L zeatin and 4% sucrose. The callus was placed in the light. As plantlets developed they were transferred to tubes containing 272K, hormone-free MS medium and 3% sucrose. The percentage of green fluorescent colonies that regenerated into whole plants was determined.

Regeneration of plants from callus transformed with PHP8080, PHP7921, and PHP8087 appeared to vary between constructs; plants were successfully regenerated from independent transformed calli at respective frequencies of 35, 54 and 86%.

Transformation was confirmed by Southern blot analyses using both the BAR and GFPm genes as hybridization probes. GFP localization in various subcellular compartments was confirmed in stable transgenic maize cells using epifluorescent microscopy and image enhancement software.

Table 3

Regeneration Frequency Of Maize Transformed With GFP
Directed to Different Subcellular Compartments

Chloroplast					
Exp #	Total Colonies	Green	Regenerate	Percentage	Ave. Plant Event
..4.01	32	23	4	17	6
..4.03	0	-	-	-	-
..4.06	13	11	8	73	3
Totals	45	34	12	35*	
Cytoplasm					
Exp #	Total Colonies	Green	Regenerate	Percentage	Ave. Plants Event
..4.01	7	6	2	33	4
..4.03	7	5	4	80	5
..4.06	8	7	6	86	4
..4.08	9	9	3	33	2
..4.11	3	3	1	33	4
Totals	34	30	16	54*	
Mitochondrial					
Exp #	Total Colonies	Green	Regenerate	Percentage	Ave. Plants Event
..4.03	5	3	1	33	9
..4.06	13	11	11	100	4
Totals	18	14	12	86*	
* This is not a mean regeneration frequency, but instead is calculated from the totals for all treatments.					

Transformed plants containing GFP in the mitochondrial subcellular compartment grew more vigorously than plants containing GFP in the cytoplasm or chloroplast. The transformed plants containing GFP in the mitochondria were equivalent to the controls in terms of growth rate and appearance.

In contrast, transformed plants containing GFP in the chloroplast were difficult to regenerate. This callus produced a preponderance of roots with few shoots. Almost all the plants containing GFP in the chloroplast that were regenerated and reached maturity clearly exhibited stress with narrow and somewhat flaccid leaves, reduced stature, yellow leaves (presumably caused by reduced chlorophyll levels) and male sterility.

Germination of T1 embryos have been tested in the following manner. Aliquots of seed from a number of independent transformants were imbibed and the embryos were excised and placed on hormone-free MS medium with 3% sucrose for germination (this in vitro test made it easier to visualize the seedling). All embryos were germinated in the dark and then scored for visual GFP expression using UV-blue excitation under the dissecting microscope.

For each event tested aliquots of 5 or 10 seeds/event were imbibed, excised and scored for germination and fluorescence. The germination frequency for excised progeny produced on plants transformed with PHP7921 and PHP8080 was good. The germination frequency for excised progeny produced on plants transformed with PHP8087 was good for 2 events (10/10), low for one event (2/10). No detectable germination was observed for 2 events. This lowered germination frequency could be due to mitochondrially-expressed GFP expression per se, but it could also be the result of event-to-event variation sometimes observed in transgenic lines.

Germination of a single event from plants transformed with PHP7921 was tested differently. Seed were screened using a UV-blue light source, and 10 seed with GFP-expressing seed (embryos and endosperm) were selected. Five of these seed were planted in soil, and in the other 5 the embryos were excised and germinated on medium. None of the seed in soil germinated, while

all excised embryos germinated, grew and the seedlings continued to fluoresce. For events where germination appears to be negatively impacted, this could be due to increased GFP concentrations during seed maturation and dry-down. High GFP concentrations and aggregation of the protein has been found to be toxic in *E. coli*. Crameri et al., *Nature Biotechnol.* 14: 315-319 (1995).

Table 4
T1 Progeny Analysis
Plant Germination and GFPm Expression

Plasmid	Event Tested	# of T1 Seed Tested	# of T1 Seed Germinated Per Event	# of T1 Plants Expressing GFPm
PHP8087: CPN60::mit-GFPm-PinII	1	10	0	0
PHP8087: CPN60::mit-GFPm-PinII	2	10	0	0
PHP8087: CPN60::mit-GFPm-PinII	3	10	10	1
PHP8087: CPN60::mit-GFPm-PinII	4	10	2	0
PHP8087: CPN60::mit-GFPm-PinII	5	10	10	2
PHP7921: Ubi::Ubi intron-GFPm-PinII	1	10	10	4
PHP8080: Ubi::Ubi intron-CTP-GFPm-PinII	1	5	5	0
PHP8080: Ubi::Ubi intron-CTP-GFPm-PinII	2	5	4	0
PHP8080: Ubi::Ubi intron-CTP-GFPm-PinII	3	5	4	0
PHP4810: Ubi::Ubi intron-BAR-PinII	1	5	5	0
*PHP7921: Ubi::Ubi intron-GFPm-PinII	2	10 5 germinated <i>in vitro</i> and 5 germinated in soil	5 Germination occurred only from <i>in vitro</i> cultures	5 Seed planted in soil also were GFPm+

* For all seed except those designated in the PHP7921 transformation, embryos were excised from imbibed seed and germinated on MS medium containing no hormones and 3% sucrose. All embryos were germinated in the dark to allow for visualization of GFPm expression. One event from PHP7921 was tested by planting one half of the seed in soil and one half with embryos excised from endosperm with culture *in vitro*. All seed planted in soil did not germinate. The germination rate for embryos excised from the endosperm and cultured *in vitro* was 100%. All plants expressed GFPm.

Example 4

Sunflower Transformation with GFP Screening, GUS Screening and Kanamycin Selection

Experiments were undertaken to test GFP, GUS and NPTII as marker genes for sunflower transformation. Two different kinds of meristem systems (split meristem; intact meristem) and two different kinds of particle systems (FG, focusing gun; He, helium gun) were used. The screenable markers GFP and GUS were tested with the intact meristem system under conditions involving no chemical selection but rather selection of transformants based on visual (GFP) or assayable (GUS) expression in transformed sectors. The NPTII marker gene was tested with the split meristem method using kanamycin for chemical selection of transformants and assays for NPTII expression to identify transformed sectors. In some cases, the experiments were carried only so far as identifying transformed sectors while other experiments were carried out all the way through to test for seed transmission of the marker gene to progeny. Transmission to progeny was confirmed by NPTII ELISA.

Regardless of the exact meristem system or type of particle gun used, the basic transformation protocol involved a combination of wounding by particle bombardment, followed by use of *Agrobacterium* for DNA delivery, as described by Bidney et al., *Plant Mol. Biol.* 18: 301-313 (1992). The plasmids used for plant transformation in these experiments were PHP167 containing GUS as the screenable marker, PHP8011 containing GFP as the screenable marker as well as the NPTII gene, and PHP762 containing NPTII as the selectable marker. Both PHP167 and PHP762 are binary vectors derived from pGA473. See An et al., *EMBO J.* 4: 277-284 (1985). The binary backbone for PHP8011 is bin 19. See Bevan, *NAR* 12: 8711-8721 (1984).

Procedures for preparation of *Agrobacterium* and preparation of particles for all of the experiments are described in Bidney et al., *supra*. The Pioneer sunflower line SMF3 was used in all experiments. See Burrus et al., *Plant Cell Rep.* 10: 161-166 (1991) for information concerning SMF3. The *Agrobacterium* strain EHA101 is described in Bidney et al. *supra* and strain EHA105 is described in Hood et al., *Transgen. Res.* 2: 208-218 (1993). Procedures for use of the helium gun, intact meristem preparation, tissue culture and co-cultivation conditions, and transgenic plant recovery for the intact meristem experiments are also described in Bidney et al., *supra*. Procedures for the use of the focusing gun, i.e., as in the intact meristem experiments, are described in Sautter et al., *Bio/Technol.* 9: 1080-1085 (1991). Procedures for use of the helium gun, split meristem preparation, tissue culture, co-cultivation, kanamycin selection and transgenic plant recovery for the split meristem experiments are described in Malone-Schoneberg et al., *Plant Sci.* 103: 199-207 (1994).

Parameters and general use of the focusing gun were similar to those 1.4 μm described in Sautter et al., *supra*. Gold particles measuring at a concentration of $0.5 \times 10^6/\mu\text{l}$ and 80 bars of nitrogen were typically used although different sized particles (0.25 to 2.5 μm) and different nitrogen pressures (40-160 bar) were tested. For the Dupont PDS-1000 helium driven particle bombardment device, the meristems were bombarded twice at the highest shelf with 600 psi rupture discs and a vacuum of 26 inches Hg and 1.8 μm tungsten particles. The intact meristem transformation method involved imbibing seed for 24 hours in the dark, removing the cotyledons and root radical, followed by culturing of the meristem explants. Twenty-four hours later, the primary leaves were removed to expose the apical meristem. The explants were placed apical dome side up and bombarded twice with particles, followed by

co-cultivation with *Agrobacterium*. To start the co-cultivation for intact meristems a 0.5 μ l droplet of OD⁶⁰⁰ 4.0 *Agrobacterium* was placed on the meristem. After a 3-day co-cultivation the meristems were transferred to culture medium with 250 μ g/ml cefotaxime (+ 100 mg/l kanamycin for the NPTII selection experiments). Selection has also been done using 50 gm/l kanamycin. The split meristem method involved imbibing seed, breaking of the cotyledons to produce a clean fracture at the plane of the embryonic axis, excising the root tip and then bisecting the explants longitudinally between the primordial leaves. The two halves were placed cut surface up on the medium then bombarded twice with particles, followed by co-cultivation with *Agrobacterium*. For split meristems, after bombardment the meristems were placed in an OD⁶⁰⁰ 0.6 *Agrobacterium* suspension for 30 minutes. They were then removed from the suspension onto solid culture medium for 3-day co-cultivation. After this period, the meristems were transferred to fresh medium with 250 μ g/ml cefotaxime (+ 100mg/l kanamycin for selection). Selection has also been done using 50 gm/l kanamycin.

Table 5 summarizes the results from experiments looking at the frequency of transformed sectors in primary shoots and secondary branches. Table 6 summarizes the results from experiments carried through to progeny for analysis of seed transmission. Transformants were identified in either seed or plants by assaying for marker gene expression. For GUS expression, activity was determined using either histochemical staining or the fluorometric assay, see Jefferson et al., *EMBO J.* 6: 3901-3907 (1987). Inheritance of transgene expression was assessed using an NPTII ELISA.

TABLE 5
Overall Transformed Sector Frequencies
for Sunflower Transformation

MARKER	PLASMID PHP #	GUN	MERISTEM PREPARATION	AVE. % SECTORS	REPS (N)
Screening:					
GUS	167	FG	Intact	22.0	15
GUS	167	FG	Intact	15.0	3
GUS	167	He	Intact	13.6	12
GFP	8011	He	Intact	7.5	1
Selection:					
NPTII/Kan	762	FG	Intact	12.7	3

TABLE 6
Overall Sector and Seed Transmission
for Sunflower Transformation

MARKER	PLASMID PHP #	GUN	MERISTEM PREPAR- ATION	AVE. % SECTORS	REPS (N)	SEED TRANS- MISSION
Screening:						
GUS	167	FG	Intact	15.0	3	0/8
GFP	8011	He	Intact	7.5	1	9/10
Selection:						
NPTII/KAN	762	FG	Intact	12.7	4	0/2
NPTII/KAN	8011	He	Split	1.8	4	6/8

For evaluation of initial sector frequencies, GUS seemed to be the most effective marker gene (Tables 5 and 6). However, based on screening alone (no selection) the observed GUS sectors did not subsequently contribute to the germline and thus did not transmit to progeny (Table 6). It was unlikely that this was due to the use of the focusing gun because, both quantitatively and qualitatively, mapping of GUS sectors after use of each type of gun was comparable. GFP was significantly better than NPTII

(7.5% vs. 1.8%, see Table 6) for both sector frequency and seed transmission when compared using the helium gun. Progeny from GFP-expressing (no chemical selection but instead visually selecting using GFP) TO plants were analyzed for NPTII expression (Table 6) Progeny from 10 plants were tested and 9 showed inheritance of NPTII activity. From kanamycin-selected TO plants, 6 out of 8 tested showed transmission of NPTII expression to the progeny. Not only has GFP "visual selection" worked unexpectedly well for recovering the primary transformants (i.e. compare Tables 3 and 4), but cotransformation and inheritance of transgene expression was also very efficient.

Table 7: Progeny Analysis of Transformation Experiments of SMF3
with GFP (PHP8011: CaMV-GFPm, ALS-NPTII)

Treatment (Comments)	SID	No. of Seeds Screened	TO NPTII Map Range	Maximum NPTII	Transmissi on Efficiency
Topdown Non-selected (from 1 experiment, 120 explants in total)	281976	246	12.6-->30	9.9	9/10
	281977	139	16.3-->	11.9	
	281978	127	>30	>30	
	281979	175	9--27	6	
	281980	74	>30	4	
	281983	15	3--21.3	6.5	
	319454	124	4.1--18.7	5.9	
	319453	118	++	8.6	
	281982	47	>30	6.2	
	281981	3	>30	0	
Split selected (from 4 experiments, 955 explants in total)	248112	6	++	2.3	6/8
	248113	24	>30	7.8	
	248111	41	>30	>30	
	248107	23	17	2.7	
	248114	67	>30	1.2	
	248110	8	4.1	0	
	248109	6	10.7	0	
	248115	86	++	5.5	

Parallel experiments comparing GFP screening, GUS screening and NPTII/kanamycin selection can be performed using intact meristems, the *Agrobacterium* strain EHA105 and the helium gun, in side-by-side comparisons. Plasmid PHP8011 can be used for GFP/NPTII comparisons. An identical plasmid substituting the GUS structural gene for GFP can be used for GUS/NPTII comparisons, and both plasmids can be used in side-by-side experiments for GUS/GFP comparisons. Sector frequency in primary shoots and secondary shoots can be

assayed and calculated as described above. Inheritance and expression of transgenes can also be analyzed in these experiments.

*Example 5**GFP As A FLP-Mediated Excision Marker In Maize*

5 Immature embryos of the Hi-II genotype were
bombarded with PHP8674 which contains a GFPm reporter
gene that is activated upon FLP recombinase-mediated
FRT excision. Plasmid PHP8674 also carries a
10 constitutively expressed gene for bialaphos resistance.
Stably transformed callus was selected on bialaphos and
examined under UV-blue light for GFP expression. No
GFP expression was observed in these primary
transformants. Pieces of callus transformed with
15 PHP8674 were then bombarded with PHP8007. Plasmid
PHP8007 contains the FLP recombinase gene operably
linked to the ubiquitin promoter. Callus sectors
expressing GFP were first observed ten days after the
second bombardment. These GFP-expressing sectors
continued to grow and express the transgene after one
20 month of culture. As a negative control the FRT/GFPm
stably transformed callus was also bombarded with a
non-FLP containing plasmid. This treatment did not
produce GFP expressing sectors.

25 As a second control for this experiment the
consecutive treatments were done in opposite order.
Hi-II immature embryos were bombarded with PHP8007
which carries the FLP recombinase gene and a bialaphos
resistance gene. Stable transformants were recovered
following selection on medium containing bialaphos.
30 These calli were re-bombarded with PHP8674. GFP
expression was observed in the callus but this faded
over 5-10 days until it could no longer be detected.
The stably transformed callus expresses FLP
recombinase. Introduction of PHP8674 likely provided
35 the substrate for excision (the FRT sites along with
spacer DNA between the sites) which activated the GFP
cassette while still on the plasmid. This "activated

expression" was only transient and disappeared as the plasmid was degraded.

Example 6**GFP Targeting to the Nucleus as a Fusion Protein**

5 Plasmid PHP9053 was constructed in which the GFP
protein was fused to the ALS enzyme (designated MALS
for maize-ALS). The ALS gene, which encodes a protein
normally found in the cytoplasm, is typically used to
10 confer herbicide resistance. Fang et al., *Plant Mol.*
Biol. **18**: 1185-1187 (1992). A nuclear localization
signal from simian virus 40 was fused to the N terminus
of GFPm as described in Example 1 (Kalderson et al.,
Cell **39**: 499-509 (1984) with the ALS enzyme fused to
15 the C terminus. The sequence encoding this nuclear
targeted fusion protein (in construct PHP9053) was
introduced into maize cells using particle bombardment
as described previously. Once gene expression was
confirmed through GFP fluorescence under the
20 microscope, tissue was fixed as described earlier and
examined to determine subcellular localization of GFP.
Fluorescence was localized to the nucleus. Plant
regeneration and assessment of toxicity for this
experiment has not yet been done.

25 GFP was targeted to the nucleus by fusing the
protein to another polypeptide that is normally
compartmentalized in the nucleus. For example, the
GFPZm-RAD51 fusion found in plasmid PHP8744 contains
RAD51, a protein required for mitotic and meiotic
30 recombination and double-stranded break repair, fused
to GFP. The gene encoding RAD51 was cloned from yeast.
Haaf et al., *Proc. Nat Acad. Sci.* **92**: 2298-2302 (1995).
Degenerate primers based on the nucleotide sequence of
the RAD51 gene cloned from yeast were used to identify
and isolate clones containing the RAD51 gene from
35 maize. The carboxy terminus of GFPm was fused to the
amino terminus of ZmRAD51a. Maize cells were
transformed with the GFPm-ZmRAD51 fusion and
fluorescence was localized in the nucleus. Plant

regeneration and assessment of toxicity for this experiment has not yet been done.

Although the foregoing refers to particular preferred embodiments, it will be understood that the present invention is not so limited. It will occur to those of ordinary skill in the art that various modifications may be made to the disclosed embodiments and that such modifications are intended to be within the scope of the present invention, which is defined by the following claims.

All publications and patent applications mentioned in this specification are indicative of the level of skill of those in the art to which the invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference in its entirety.

What Is Claimed Is:

1. An isolated DNA molecule comprising a nucleotide sequence encoding a green fluorescent protein operably linked to an inducible promoter.

2. The isolated DNA sequence according to claim 1, wherein the inducible promoter is selected from the group consisting of the estrogen-inducible promoter, the estradiol-inducible promoter, the ACE1 promoter, the IN2 promoter and the tetracycline repressor promoter.

3. An isolated DNA molecule according to claim 1, wherein said nucleotide sequence encoding a green fluorescent protein is operably linked to a nucleotide sequence encoding a signal sequence for subcellular localization which directs a protein to a subcellular compartment.

4. An isolated DNA molecule comprising a nucleotide sequence selected from the group consisting of:

- (a) SEQ ID NO: 1;
- (b) a nucleotide sequence that has substantial sequence similarity with SEQ ID NO: 1; and
- (c) a functional fragment of (a) or (b), wherein said DNA molecule encodes a green fluorescent protein.

5. An isolated DNA molecule according to claim 4, wherein said nucleotide sequence encoding a green fluorescent protein is operably linked to a nucleotide sequence encoding a signal sequence for subcellular localization which directs a protein to a subcellular compartment.

6. An isolated DNA molecule according to claim 5, further comprising a promoter operably linked to said nucleotide sequence encoding a signal sequence and a green fluorescent protein.

7. An expression vector comprising said DNA molecule of claim 1.

8. An expression vector comprising said DNA molecule of claim 4.

9. The expression vector according to claim 8, wherein said nucleotide sequence encoding a green fluorescent protein is operably linked to a nucleotide sequence encoding a signal sequence which directs a protein to a subcellular compartment.

10. The expression vector according to claim 9, further comprising a first promoter operably linked to said nucleotide sequence encoding a signal sequence and a green fluorescent protein.

11. The expression vector according to claim 10, further comprising a foreign gene, wherein said foreign gene is operably linked to a second promoter.

12. A method of using said expression vector of claim 7 to produce a transformed plant, comprising the steps of introducing said DNA molecule into regenerable plant cells, inducing expression of the gene encoding the green fluorescent protein, and selecting cells containing said green fluorescent protein for regeneration.

13. A method of using said expression vector of claim 8 to produce a transformed plant, comprising the steps of introducing said expression vector into

regenerable plant cells and selecting cells containing said green fluorescent protein for regeneration.

14. The method of claim 12, wherein said regenerable plant cells are selected from the group consisting of *Zea*, *Brassica* and *Helianthus* cells.

15. The method of claim 13, wherein said regenerable plant cells are selected from the group consisting of *Zea*, *Brassica* and *Helianthus* cells.

16. A transgenic plant expressing the isolated DNA molecule of claim 1.

17. A transgenic plant comprising the vector of claim 4.

18. A method for producing a transgenic plant, comprising the steps:

- (a) constructing an expression vector comprising (i) a first promoter which is an inducible promoter, operably linked to a nucleotide sequence encoding a green fluorescent protein, and (ii) a second promoter operably linked to a foreign gene;
- (b) introducing said expression vector into regenerable plant cells;
- (c) inducing expression of the gene encoding the green fluorescent protein and selecting transformed plant cells containing said protein; and
- (d) regenerating transformed plants from said selected transformed plant cells.

19. The method of claim 18, where said inducible promoter is selected from the group consisting of the estrogen-inducible promoter, the

estradiol-inducible promoter, the ACE1 promoter, the IN2 promoter and the tetracycline repressor promoter.

20. A method for producing a transgenic plant, comprising the steps:

- (a) constructing an expression vector comprising (i) a first promoter operably linked to a nucleotide sequence encoding a sequence for subcellular localization which directs a protein to a subcellular compartment and nucleotide sequence encoding a green fluorescent protein, and (ii) a second promoter operably linked to a foreign gene;
- (b) introducing said expression vector into regenerable plant cells;
- (c) selecting transformed plant cells containing said green fluorescent protein; and
- (d) regenerating transformed plants from said selected transformed plant cells.

21. The method of claim 20, wherein the sequence for subcellular localization is selected from the group which through targeting, signaling and/or retention localize the encoded protein to mitochondria, chloroplasts, peroxisomes, vacuole, endoplasmic reticulum, cell wall or for secretion generally into the apoplast.

22. The method according to claim 18, wherein said nucleotide sequence encoding a green fluorescent protein is selected from the group consisting of:

- (a) SEQ ID NO: 1;
- (b) a nucleotide sequence that has substantial sequence similarity with SEQ ID NO: 1; and

- (c) a functional fragment of (a) or (b).

23. A method for producing a transgenic plant, comprising the steps:

- (a) constructing an expression vector comprising (i) a first promoter operably linked to a nucleotide sequence encoding a screenable marker flanked on the 5-prime and 3-prime ends with a recombinase-specific target sequence, and (ii) a second promoter operably linked to a foreign gene;
- (b) introducing said expression vector into regenerable plant cells;
- (c) selecting transformed plant cells containing said screenable marker;
- (d) transforming the plant cells with a second expression vector containing a gene encoding a site-specific recombinase;
- (e) selecting plant cells that no longer express the screenable marker;
- (f) regenerating transformed plants from said selected transformed plant cells; and
- (g) isolating said foreign protein.

24. The method according to claim 23, wherein said site-specific recombinase is selected from the group consisting of the FLP/FRT, Ac/DS and cre/lox systems.

25. The method according to claim 23, wherein said screenable marker is the green fluorescent protein.

26. An isolated DNA molecule comprising a nucleotide sequence encoding the green fluorescent

protein fused in frame to superoxide dismutase such that cells transformed with said DNA molecule contain a fusion protein that (i) produces green fluorescence in the presence of UV to blue light and (ii) displays superoxide dismutase activity.

27. A method for producing a transgenic plant comprising:

- (a) constructing an expression vector comprising (i) a first promoter operably linked to a nucleotide sequence encoding a green fluorescent protein, and (ii) a second promoter operably linked to a gene encoding an enzyme that is an oxygen scavenger, and (iii) a third promoter operably linked to a foreign gene;
- (b) introducing said expression vector into regenerable plant cells;
- (c) selecting transformed plant cells containing said green fluorescent protein; and
- (d) regenerating transformed plants from said selected transformed plant cells.

28. The method according to claim 27, wherein said oxygen scavenger enzyme is superoxide dismutase.

29. A method for producing a transgenic plant comprising:

- (a) constructing an expression vector comprising (i) a first promoter operably linked to a nucleotide sequence encoding a fusion protein comprising the green fluorescent protein and an oxygen scavenger enzyme fused in frame, and

- (ii) a second promoter operably linked to a foreign gene;
- (b) introducing said expression vector into regenerable plant cells;
- (c) selecting transformed plant cells containing said green fluorescent protein and oxygen scavenger enzyme activity; and
- (d) regenerating transformed plants from said selected transformed plant cells.

30. The method according to claim 29, wherein said oxygen scavenger enzyme is superoxide dismutase.

1/5

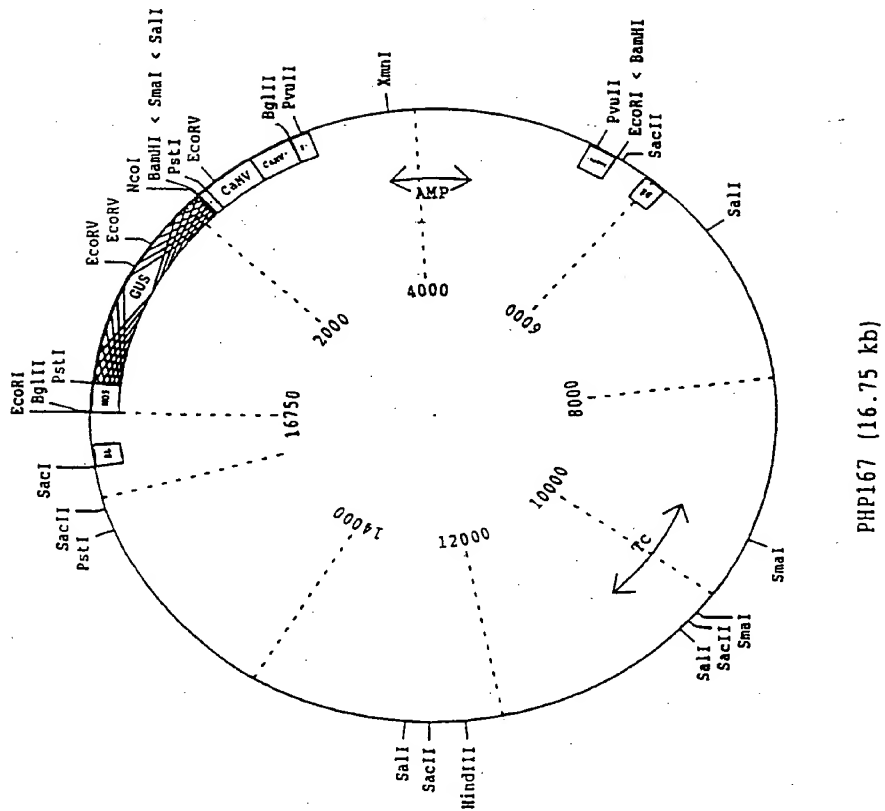
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61  GAGGTGAACGGGCCACAAGTTCTCCGTGTCCGGCGAGGGCGAGGGCGACCCACCTACCGG
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121  AAGCTCACCCCTCAAGTTCATCTGCACCAACGGCAAGCTCCCGGTGCCCTGGCCCAACCTC
-----> 180
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-----> 240
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361  AACAGGATTGAGCTCAAGGGCATCGACTTCAAGGAGGACGGCAACATCTCTGGCCCAAG
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481  ATCAAGGTGAACCTCAAGATCAGGCACAACATCGAGGACGGCTCACTGCGAGCTCGTGCAC
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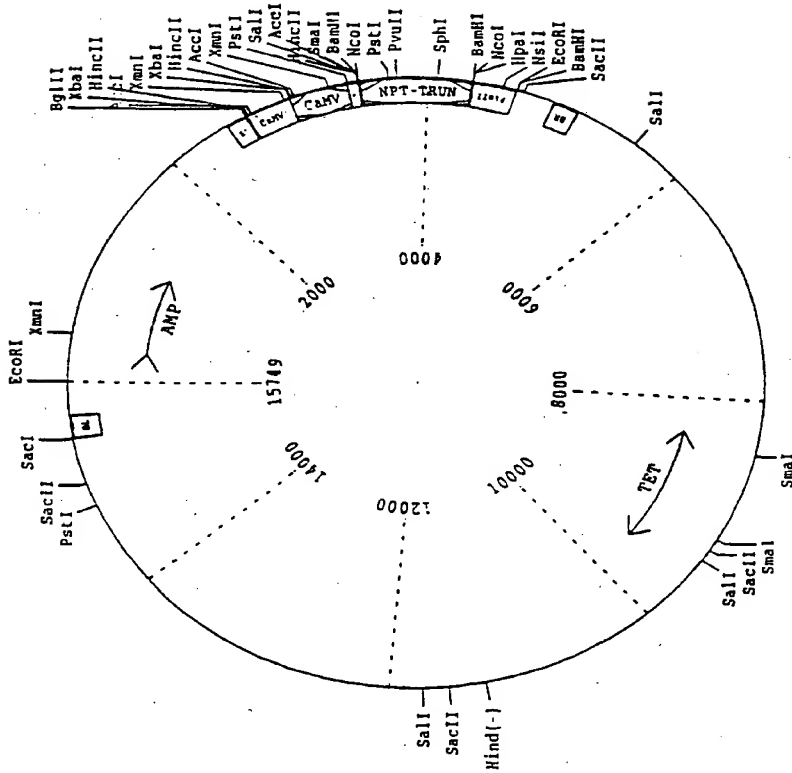
FIGURE 1



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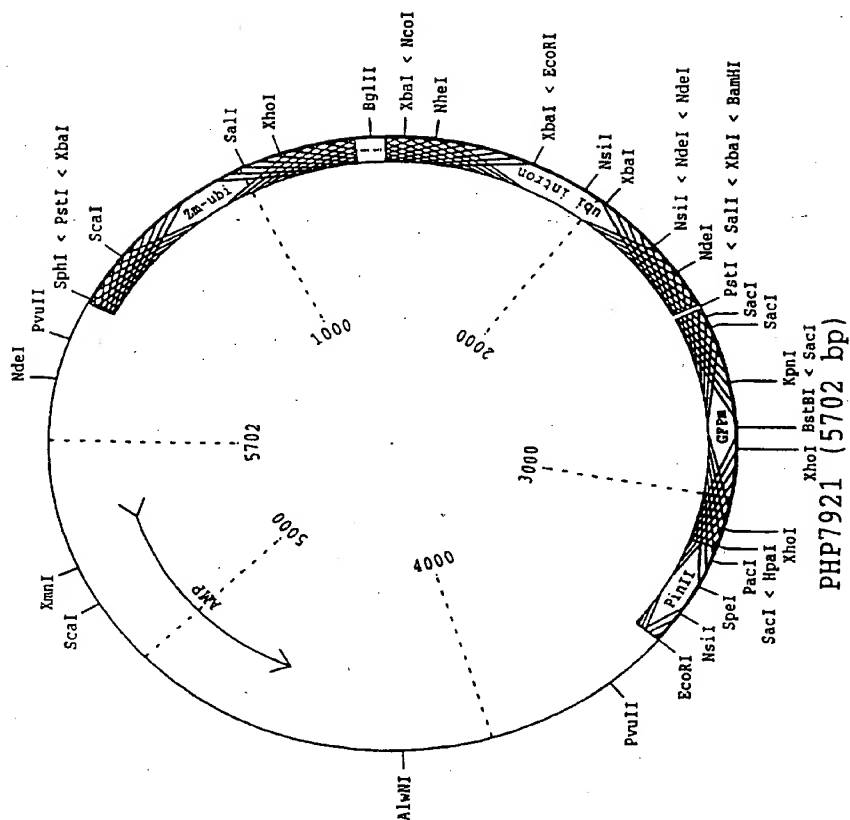
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FIGURE 2



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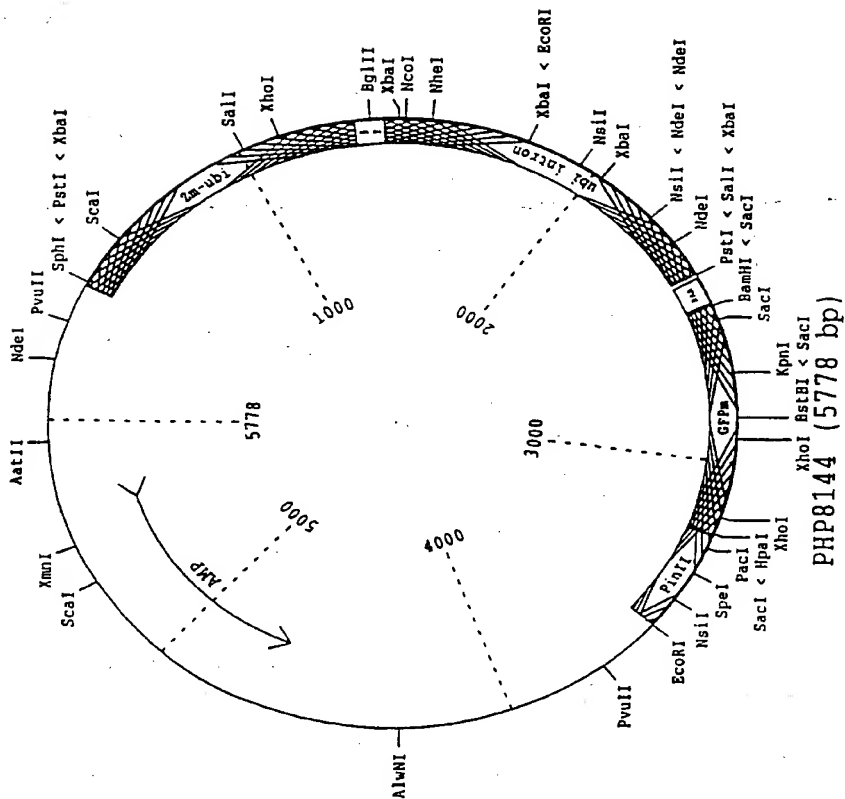
FIGURE 3



PLASMIDMAP of: Ubigfpm.Seq check: 822 from: 1 to: 5702

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FIGURE 4



PLASMIDMAP of PHP8144. Seq check: 8061 from: 1 to: 5778

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FIGURE 5